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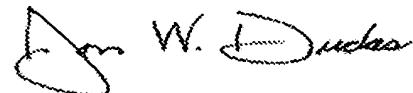
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APPLICATION NUMBER: 60/560,147

FILING DATE: *April 07, 2004*

RELATED PCT APPLICATION NUMBER: PCT/US04/29047

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MAIL STOP PROVISIONAL PATENT APPLICATION
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CERTIFICATE OF EXPRESS MAILING

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I hereby certify that this correspondence is being deposited with the United States Postal Service as "Express Mail Post Office to Address" service under 37 CFR 1.10 on the date indicated above and is addressed to: Mail Stop Provisional Patent Application, Commissioner for Patents, P.O. Box 1450, Alexandria, VA 22313-1450

Kathy Hart Gagnon
Kathy Hart Gagnon

PROVISIONAL APPLICATION TRANSMITTAL

(REQUEST FOR FILING A PROVISIONAL APPLICATION FOR PATENT UNDER 37 CFR § 1.53(C))

Dear Sir:

Please find enclosed a provisional patent application and papers as follows for:

Samuel J.	DANISHEFSKY	22 Brayton Street Englewood, NJ 07631
J. David	WARREN	1233 York Avenue, Apt 3E New York, New York 10021

Title of the Invention: METHOD FOR PREPARING POLYFUNCTIONALIZED PEPTIDES AND/OR PROTEINS VIA NATIVE CHEMICAL LIGATION

A) ENCLOSED APPLICATION PARTS:

1) <input checked="" type="checkbox"/> Specification, consisting of:	PAGES	<u>56</u>
a) <input checked="" type="checkbox"/> Description (55 pages)		
b) <input checked="" type="checkbox"/> Abstract (1 page)		
2) <input checked="" type="checkbox"/> Drawings	SHEETS	<u>2</u>
	TOTAL	<u>58</u>

B) OTHER ACCOMPANYING APPLICATION PARTS:

- 3) Return Receipt Postcard (MPEP § 503) (specifically itemized)
- 4) Application Data Sheet. See 37 CFR § 1.76
- 5) OTHER: (if applicable, specified below)

C) CORRESPONDENCE ADDRESS:

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D) METHOD OF PAYMENT OF FILING FEES:

Applicant claims small entity status. See 37 CFR §1.27.

Statement Verifying Small Entity Status (optional)

A check or money order is enclosed to cover the filing fees.

The Commissioner is hereby authorized to charge filing fees or credit any overpayment to Deposit Account Number: 03-1721.

FILING FEE AMOUNT (\$): \$ 80.00

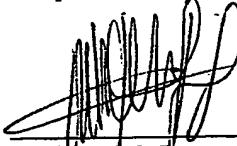
E) GOVERNMENT SUPPORT

THE INVENTION WAS MADE, IN PART, BY AN AGENCY OF THE UNITED STATES GOVERNMENT
OR UNDER A CONTRACT WITH AN AGENCY OF THE UNITED STATES GOVERNMENT:

 NO.

YES, THE NAME OF THE U.S. GOVERNMENT AGENCY AND THE GOVERNMENT
CONTRACT NUMBER ARE : GRANT NOS.: CA103823 (FORMERLY AI16943), CA62948
AND AI051883 FROM THE NATIONAL INSTITUTES OF HEALTH; AND PC020147 FROM
THE U.S. ARMY PROSTATE CANCER RESEARCH PROGRAM.

Respectfully Submitted,



Nadege M. Lagneau, Ph.D.
Reg. No. 51,908

Dated: April 7, 2004

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METHOD FOR PREPARING POLYFUNCTIONALIZED PEPTIDES AND/OR PROTEINS VIA NATIVE CHEMICAL LIGATION

RELATED APPLICATIONS

[0001] This application is related to provisional application No.: 60/500,708, filed September 5, 2003; which is hereby incorporated by reference in its entirety.

GOVERNMENT SUPPORT

[0002] The invention was supported in part by Grant Nos.: CA103823 (formerly AI16943), CA62948 and AI051883 from the National Institutes of Health; and PC020147 from the U.S. Army Prostate Cancer Research Program. The U.S. government may have certain rights in this invention.

BACKGROUND OF THE INVENTION

[0003] Glycoproteins are important biomacromolecules that are biosynthesized through posttranslational glycosylation of newly fashioned proteins emerging from the ribosome. Interest in glycoproteins arises at many levels. A long-term goal of the growing field of chemistry-based glycobiology is the delineation of the consequences of glycosylation on critical properties such as protein folding, proteolytic stability, and cell adhesion.¹ Such insights could explain why nature bothers to glycosylate otherwise functional proteins. Moreover, glycoproteins have potentially important clinical roles in the contexts of vaccines, diagnostics, and therapeutics. Indeed, erythropoietin (1), albeit a heterogeneous glycoprotein,⁵ is clinically valuable as a treatment for anemia, among other indications.^{4a}

[0004] Many naturally occurring, medicinally important glycoproteins (*cf.*, for example, erythropoietin^{4a} and gp120¹¹) display multiple glycosylation sites containing large oligosaccharide domains. However, given the complexity and variability of biological glycosylation pathways,¹⁰ the isolation of homogeneous glycoproteins from natural sources in significant quantity is extremely difficult.

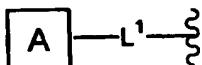
[0005] Numerous methods exist for the production of glycopeptides by chemical synthesis. For example, glycans have been introduced into peptides *via* amino acid "cassettes" with pendant protected saccharides,²⁶⁻³¹ through enzymatic manipulations of glycopeptides,³²⁻³⁸

or by conjugation of fully elaborated, complex saccharides to short synthetic peptides.³⁹⁻⁴¹ Larger *O*-linked glycopeptides have been synthesized using ligation techniques^{42,43} such as expressed protein ligation.⁴⁴⁻⁴⁶ Bertozzi and coworkers extended the scope of the "cassette" approach by applying native chemical ligation to the synthesis of a biologically active glycoprotein with two single-residue *O*-linked glycans.⁴⁷ Tolbert and Wong described the ligation of a 392-residue intein-generated peptide thioester and a dipeptide functionalized with a single *N*-acetylglucosamine residue. However, none of these approaches has allowed the assembly of complex glyopeptides or glycoproteins multiply functionalized (e.g., multiply glycosylated) at designated sites.

[0006] Accordingly, there remains a need for novel synthetic methods leading to the preparation of homogeneous polyfunctionalized peptides and/or proteins. Specifically, convergent, stereoselective, versatile methods for preparing such glycopeptides and/or glycoproteins are needed.

SUMMARY OF THE INVENTION

[0007] In recognition of the need to provide access to polyfunctionalized peptides and proteins, the present invention, in one aspect, provides a system for preparing a polyfunctionalized peptide comprising a peptidic backbone made up of four or more amino acids, wherein two or more non-adjacent amino acids are independently substituted with a moiety having the structure:



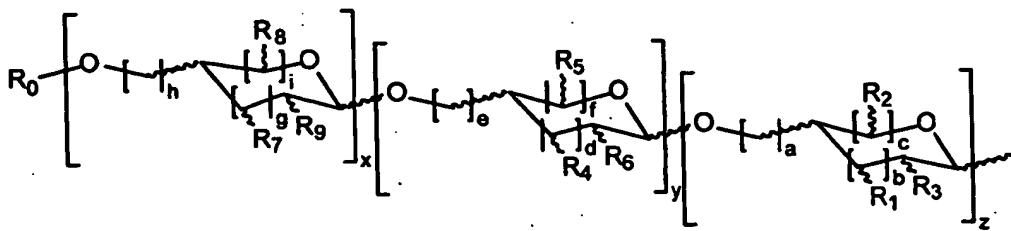
wherein each occurrence of L¹ is independently a substituted or unsubstituted, linear or branched, cyclic or acyclic, saturated or unsaturated aliphatic or heteroaliphatic moiety;

each occurrence of A is independently an aliphatic, heteroaliphatic, aryl, heteroaryl or a pharmaceutically useful group or entity;

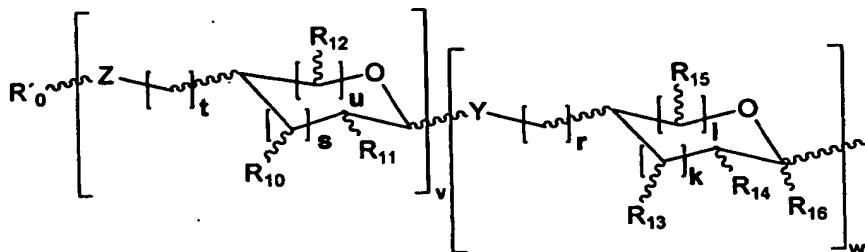
with the proviso that the peptide sequence between any two consecutive, non-adjacent, amino acids bearing a A-L¹- moiety comprises at least one cysteine residue.

[0008] In certain embodiments, each occurrence of A is independently a pharmaceutically useful group or entity. In certain embodiments, each occurrence of A is independently a biomolecule, a small molecule, a macromolecule or a diagnostic label.

[0009] In certain exemplary embodiments, each occurrence of A is independently a carbohydrate determinant having the structure:



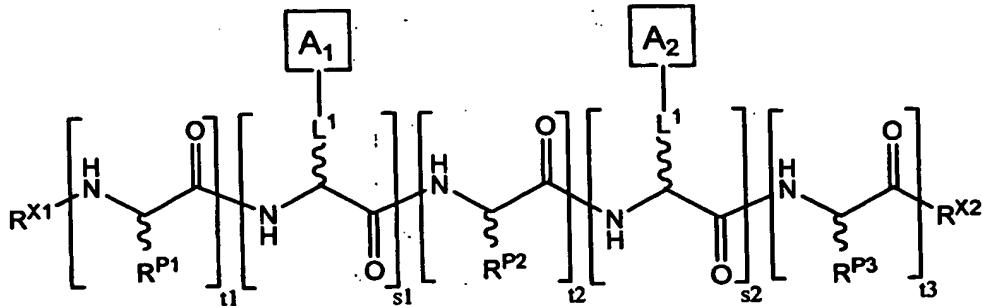
wherein a, b, c, d, e, f, g, h, i, x, y and z are independently 0, 1, 2 or 3, with the proviso that the x, y and z bracketed structures represent furanose or pyranose moieties and the sum of b and c is 1 or 2, the sum of d and f is 1 or 2, and the sum of g and i is 1 or 2, and with the proviso that x, y and z are not simultaneously 0; wherein R₀ is hydrogen, a linear or branched chain alkyl, acyl, arylalkyl or aryl group; wherein each occurrence of R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈ and R₉ is independently hydrogen, OH, ORⁱ, NHRⁱ, NHCORⁱ, F, CH₂OH, CH₂ORⁱ, a substituted or unsubstituted linear or branched chain alkyl, (mono-, di- or tri)hydroxyalkyl, (mono-, di- or tri)acyloxyalkyl, arylalkyl or aryl group; wherein each occurrence of Rⁱ is independently hydrogen, CHO, COORⁱⁱ, or a substituted or unsubstituted linear or branched chain alkyl, acyl, arylalkyl or aryl group or a saccharide moiety having the structure:



wherein Y and Z are independently NH or O; wherein k, l, r, s, t, u, v and w are each independently 0, 1 or 2; with the proviso that the v and w bracketed structures represent furanose or pyranose moieties and the sum of l and k is 1 or 2, and the sum of s and u is 1 or 2, and with the proviso that v and w are not simultaneously 0; wherein R'0 is hydrogen, a linear or branched chain alkyl, acyl, arylalkyl or aryl group; wherein each occurrence of R₁₀, R₁₁, R₁₂, R₁₃, R₁₄ and R₁₅ is independently hydrogen, OH, ORⁱⁱⁱ, NHRⁱⁱⁱ, NHCORⁱⁱⁱ, F, CH₂OH, CH₂ORⁱⁱⁱ, or a substituted or unsubstituted linear or branched chain alkyl, (mono-, di- or tri)hydroxyalkyl, (mono-, di- or tri)acyloxyalkyl, arylalkyl or aryl group; wherein each occurrence of R₁₆ is

hydrogen, COOH, COORⁱⁱ, CONHRⁱⁱ, a substituted or unsubstituted linear or branched chain alkyl or aryl group; wherein each occurrence of Rⁱⁱⁱ is hydrogen, CHO, COOR^{iv}, or a substituted or unsubstituted linear or branched chain alkyl, acyl, arylalkyl or aryl group; and wherein each occurrence of Rⁱⁱ and R^{iv} are each independently H, or a substituted or unsubstituted linear or branched chain alkyl, arylalkyl or aryl group.

[0010] In certain embodiments, the present invention provides a method for preparing a polyfunctionalized peptide having the structure:



s1 and s2 are independently an integer from 1 to about 20;

t1, t2 and t3 are each independently an integer;

R^{X1} is hydrogen, alkyl, acyl, aryl, heteroaryl, -alkyl(aryl), -alkyl(heteroaryl), a nitrogen protecting group, an amino acid or a protected amino acid;

R^{X2} is -OR^{X2a} or -NR^{X2b}R^{X2c}, wherein R^{X2a} is hydrogen, alkyl, aryl, heteroaryl, -alkyl(aryl), -alkyl(heteroaryl), a carboxylic acid protecting group, an amino acid or a protected amino acid; and R^{X2b} and R^{X2c} are independently hydrogen, alkyl, aryl, heteroaryl, -alkyl(aryl), -alkyl(heteroaryl), a nitrogen protecting group, an amino acid or a protected amino acid;

R^{P1}, R^{P2} and R^{P3} are independently H, alkyl, heteroalkyl, aryl, heteroaryl, -alkyl(aryl), -alkyl(heteroaryl), or a natural or non-natural amino acid side chain;

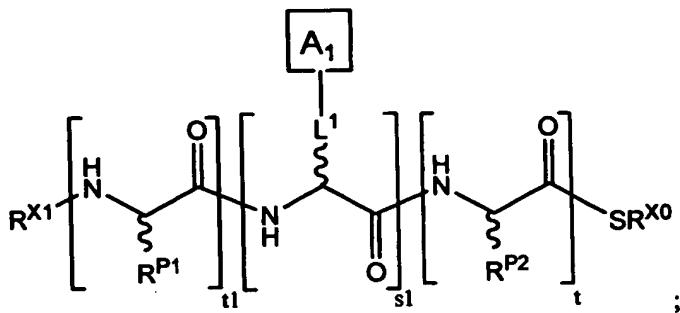
each occurrence of L¹ is independently a substituted or unsubstituted aliphatic or heteroaliphatic moiety;

A₁ and A₂ are each independently an aliphatic, heteroaliphatic, aryl, heteroaryl or a pharmaceutically useful group or entity; and

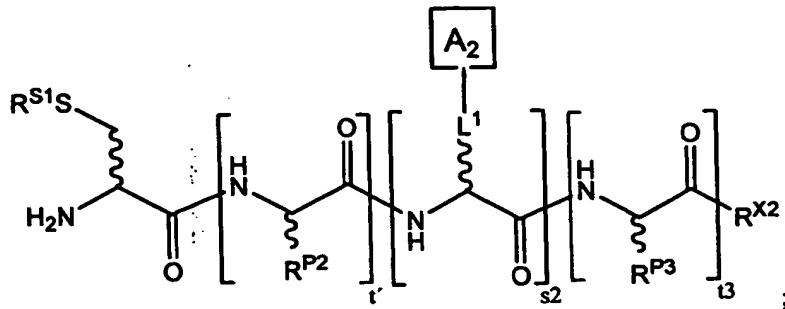
at least one occurrence of the bracketed structure t2 is a cysteine residue or protected cysteine residue;

wherein the method comprise a step of:

reacting a peptide acyl donor having the structure:



with a peptide amine acceptor having the structure:

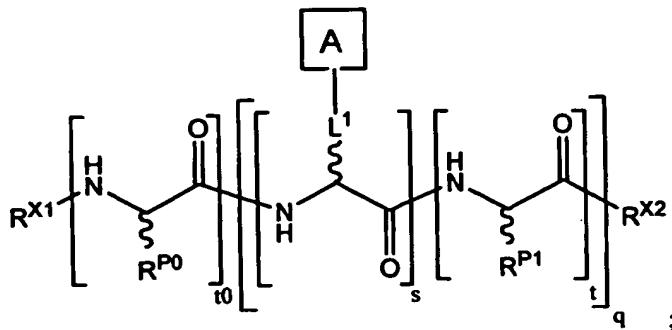


under suitable conditions to effect disulfide exchange ligation;

wherein the sum $t+t'$ equals $(t_2)+1$; R^{S_1} is a sulfide protecting group; and R^{X_0} is a group such that the moiety $-C(=O)SR^{X_0}$ can be made to undergo disulfide exchange with the glycopeptide amine acceptor.

[0011] In certain exemplary embodiments, A_1 and A_2 are each independently a carbohydrate domain as defined for A above.

[0012] In certain embodiments, the step may be carried out once, or repeated a desired number of times, to prepare the polyfunctionalized peptide having the structure:



wherein R^{X_1} and R^{X_2} are as defined above;

each occurrence of A may be the same or different and may be as defined for A₁ and A₂ above;

each occurrence of R^{P1} may be the same or different and may be as defined for R^{P1} and R^{P2} above;

q is an integer greater than or equal to 2;

each occurrence of s is independently an integer from 1 to about 20;

each occurrence of t is independently an integer;

t0 is an integer; and

each occurrence of R^{P0} is independently H, alkyl, heteroalkyl, aryl, heteroaryl, -alkyl(aryl), -alkyl(heteroaryl), or a natural or non-natural amino acid side chain.

DEFINITIONS

[0013] Certain specific functional groups defined in the inventive method are described in more detail below. For purposes of this invention, the chemical elements are identified in accordance with the Periodic Table of the Elements, CAS version, Handbook of Chemistry and Physics, 75th Ed., inside cover, and specific functional groups are defined as described therein. Additionally, general principles of organic chemistry, as well as specific functional moieties and reactivity, are described in "Organic Chemistry", Thomas Sorrell, University Science Books, Sausalito: 1999, the entire contents of which are incorporated herein by reference.

[0014] It will be appreciated that the compounds, as described herein, may be substituted with any number of substituents or functional moieties. In general, the term "substituted" whether preceded by the term "optionally" or not, and substituents contained in formulas of this invention, refer to the replacement of hydrogen radicals in a given structure with the radical of a specified substituent. When more than one position in any given structure may be substituted with more than one substituent selected from a specified group, the substituent may be either the same or different at every position unless otherwise indicated. As used herein, the term "substituted" is contemplated to include all permissible substituents of organic compounds. In a broad aspect, the permissible substituents include acyclic and cyclic, branched and unbranched, carbocyclic and heterocyclic, aromatic and nonaromatic substituents of organic compounds. For purposes of this invention, heteroatoms such as nitrogen may have hydrogen substituents and/or any permissible substituents of organic compounds described herein which satisfy the valencies

of the heteroatoms. Furthermore, this invention is not intended to be limited in any manner by the permissible substituents of organic compounds. Combinations of substituents and variables envisioned by this invention are preferably those that result in the formation of stable compounds useful in the treatment and/or prevention of HIV, or in the inducement of antibodies, as described herein. The term "stable", as used herein, preferably refers to compounds which possess stability sufficient to allow manufacture and which maintain the integrity of the compound for a sufficient period of time to be useful for the purposes detailed herein.

[0015] The term "aliphatic", as used herein, includes both saturated and unsaturated, straight chain (*i.e.*, unbranched) or branched aliphatic hydrocarbons, which are optionally substituted with one or more functional groups. As will be appreciated by one of ordinary skill in the art, "aliphatic" is intended herein to include, but is not limited to, alkyl, alkenyl, alkynyl moieties. Thus, as used herein, the term "alkyl" includes straight and branched alkyl groups. An analogous convention applies to other generic terms such as "alkenyl", "alkynyl" and the like. Furthermore, as used herein, the terms "alkyl", "alkenyl", "alkynyl" and the like encompass both substituted and unsubstituted groups. In certain embodiments, as used herein, "lower alkyl" is used to indicate those alkyl groups (cyclic, acyclic, substituted, unsubstituted, branched or unbranched) having 1-6 carbon atoms.

[0016] In certain embodiments, the alkyl, alkenyl and alkynyl groups employed in the invention contain 1-20 aliphatic carbon atoms. In certain other embodiments, the alkyl, alkenyl, and alkynyl groups employed in the invention contain 1-10 aliphatic carbon atoms. In yet other embodiments, the alkyl, alkenyl, and alkynyl groups employed in the invention contain 1-8 aliphatic carbon atoms. In still other embodiments, the alkyl, alkenyl, and alkynyl groups employed in the invention contain 1-6 aliphatic carbon atoms. In yet other embodiments, the alkyl, alkenyl, and alkynyl groups employed in the invention contain 1-4 carbon atoms. Illustrative aliphatic groups thus include, but are not limited to, for example, methyl, ethyl, n-propyl, isopropyl, allyl, n-butyl, sec-butyl, isobutyl, tert-butyl, n-pentyl, sec-pentyl, isopentyl, tert-pentyl, n-hexyl, sec-hexyl, moieties and the like, which again, may bear one or more substituents. Alkenyl groups include, but are not limited to, for example, ethenyl, propenyl, butenyl, 1-methyl-2-buten-1-yl, and the like. Representative alkynyl groups include, but are not limited to, ethynyl, 2-propynyl (propargyl), 1-propynyl and the like.

[0017] The term "alicyclic", as used herein, refers to compounds which combine the properties of aliphatic and cyclic compounds and include but are not limited to cyclic, or polycyclic aliphatic hydrocarbons and bridged cycloalkyl compounds, which are optionally substituted with one or more functional groups. As will be appreciated by one of ordinary skill in the art, "alicyclic" is intended herein to include, but is not limited to, cycloalkyl, cycloalkenyl, and cycloalkynyl moieties, which are optionally substituted with one or more functional groups. Illustrative alicyclic groups thus include, but are not limited to, for example, cyclopropyl, -CH₂-cyclopropyl, cyclobutyl, -CH₂-cyclobutyl, cyclopentyl, -CH₂-cyclopentyl-n, cyclohexyl, -CH₂-cyclohexyl, cyclohexenylethyl, cyclohexanylethyl, norborbyl moieties and the like, which again, may bear one or more substituents.

[0018] The term "alkoxy" (or "alkyloxy"), or "thioalkyl" as used herein refers to an alkyl group, as previously defined, attached to the parent molecular moiety through an oxygen atom or through a sulfur atom. In certain embodiments, the alkyl group contains 1-20 aliphatic carbon atoms. In certain other embodiments, the alkyl group contains 1-10 aliphatic carbon atoms. In yet other embodiments, the alkyl, alkenyl, and alkynyl groups employed in the invention contain 1-8 aliphatic carbon atoms. In still other embodiments, the alkyl group contains 1-6 aliphatic carbon atoms. In yet other embodiments, the alkyl group contains 1-4 aliphatic carbon atoms. Examples of alkoxy, include but are not limited to, methoxy, ethoxy, propoxy, isopropoxy, n-butoxy, tert-butoxy, neopentoxy and n-hexoxy. Examples of thioalkyl include, but are not limited to, methylthio, ethylthio, propylthio, isopropylthio, n-butylthio, and the like.

[0019] The term "alkylamino" refers to a group having the structure -NHR' wherein R' is alkyl, as defined herein. The term "aminoalkyl" refers to a group having the structure NH₂R', wherein R' is alkyl, as defined herein. In certain embodiments, the alkyl group contains 1-20 aliphatic carbon atoms. In certain other embodiments, the alkyl group contains 1-10 aliphatic carbon atoms. In yet other embodiments, the alkyl, alkenyl, and alkynyl groups employed in the invention contain 1-8 aliphatic carbon atoms. In still other embodiments, the alkyl group contains 1-6 aliphatic carbon atoms. In yet other embodiments, the alkyl group contains 1-4 aliphatic carbon atoms. Examples of alkylamino include, but are not limited to, methylamino, ethylamino, iso-propylamino and the like.

[0020] Some examples of substituents of the above-described aliphatic (and other) moieties of compounds of the invention include, but are not limited to aliphatic; heteroaliphatic;

aryl; heteroaryl; alkylaryl; alkylheteroaryl; alkoxy; aryloxy; heteroalkoxy; heteroaryloxy; alkylthio; arylthio; heteroalkylthio; heteroarylthio; F; Cl; Br; I; -OH; -NO₂; -CN; -CF₃; -CH₂CF₃; -CHCl₂; -CH₂OH; -CH₂CH₂OH; -CH₂NH₂; -CH₂SO₂CH₃; -C(O)R_x; -CO₂(R_x); -CON(R_x)₂; -OC(O)R_x; -OCO₂R_x; -OCON(R_x)₂; -N(R_x)₂; -S(O)₂R_x; -NR_x(CO)R_x wherein each occurrence of R_x independently includes, but is not limited to, aliphatic, heteroaliphatic, aryl, heteroaryl, alkylaryl, or alkylheteroaryl, wherein any of the aliphatic, heteroaliphatic, alkylaryl, or alkylheteroaryl substituents described above and herein may be substituted or unsubstituted, branched or unbranched, cyclic or acyclic, and wherein any of the aryl or heteroaryl substituents described above and herein may be substituted or unsubstituted. Additional examples of generally applicable substituents are illustrated by the specific embodiments shown in the Examples that are described herein.

[0021] In general, the terms "aryl" and "heteroaryl", as used herein, refer to stable mono- or polycyclic, heterocyclic, polycyclic, and polyheterocyclic unsaturated moieties having preferably 3-14 carbon atoms, each of which may be substituted or unsubstituted. It will also be appreciated that aryl and heteroaryl moieties, as defined herein may be attached via an aliphatic, alicyclic, heteroaliphatic, heteroalicyclic, alkyl or heteroalkyl moiety and thus also include -(aliphatic)aryl, -(heteroaliphatic)aryl, -(aliphatic)heteroaryl, -(heteroaliphatic)heteroaryl, -(alkyl)aryl, -(heteroalkyl)aryl, -(heteroalkyl)aryl, and -(heteroalkyl)heteroaryl moieties. Thus, as used herein, the phrases "aryl or heteroaryl" and "aryl, heteroaryl, -(aliphatic)aryl, -(heteroaliphatic)aryl, -(aliphatic)heteroaryl, -(heteroaliphatic)heteroaryl, -(alkyl)aryl, -(heteroalkyl)aryl, -(heteroalkyl)aryl, and -(heteroalkyl)heteroaryl" are interchangeable. Substituents include, but are not limited to, any of the previously mentioned substituents, *i.e.*, the substituents recited for aliphatic moieties, or for other moieties as disclosed herein, resulting in the formation of a stable compound. In certain embodiments of the present invention, "aryl" refers to a mono- or bicyclic carbocyclic ring system having one or two aromatic rings including, but not limited to, phenyl, naphthyl, tetrahydronaphthyl, indanyl, indenyl and the like. In certain embodiments of the present invention, the term "heteroaryl", as used herein, refers to a cyclic aromatic radical having from five to ten ring atoms of which one ring atom is selected from S, O and N; zero, one or two ring atoms are additional heteroatoms independently selected from S, O and N; and the remaining ring atoms are carbon, the radical being joined to the rest of the molecule via any of the ring atoms, such as, for example, pyridyl, pyrazinyl, pyrimidinyl,

pyrrolyl, pyrazolyl, imidazolyl, thiazolyl, oxazolyl, isooxazolyl, thiadiazolyl, oxadiazolyl, thiophenyl, furanyl, quinolinyl, isoquinolinyl, and the like.

[0022] It will be appreciated that aryl and heteroaryl groups (including bicyclic aryl groups) can be unsubstituted or substituted, wherein substitution includes replacement of one, two or three of the hydrogen atoms thereon independently with any one or more of the following moieties including, but not limited to: aliphatic; heteroaliphatic; aryl; heteroaryl; alkylaryl; alkylheteroaryl; alkoxy; aryloxy; heteroalkoxy; heteroaryloxy; alkylthio; arylthio; heteroalkylthio; heteroarylthio; F; Cl; Br; I; -OH; -NO₂; -CN; -CF₃; -CH₂CF₃; -CHC1₂; -CH₂OH; -CH₂CH₂OH; -CH₂NH₂; -CH₂SO₂CH₃; -C(O)R_x; -CO₂(R_x); -CON(R_x)₂; -OC(O)R_x; -OCO₂R_x; -OCON(R_x)₂; -N(R_x)₂; -S(O)₂R_x; -NR_x(CO)R_x wherein each occurrence of R_x independently includes, but is not limited to, aliphatic, heteroaliphatic, aryl, heteroaryl, alkylaryl, or alkylheteroaryl, wherein any of the aliphatic, heteroaliphatic, alkylaryl, or alkylheteroaryl substituents described above and herein may be substituted or unsubstituted, branched or unbranched, cyclic or acyclic, and wherein any of the aryl or heteroaryl substituents described above and herein may be substituted or unsubstituted. Additional examples of generally applicable substituents are illustrated by the specific embodiments shown in the Examples that are described herein.

[0023] The term "cycloalkyl", as used herein, refers specifically to groups having three to seven, preferably three to ten carbon atoms. Suitable cycloalkyls include, but are not limited to cyclopropyl, cyclobutyl, cyclopentyl, cyclohexyl, cycloheptyl and the like, which, as in the case of aliphatic, heteroaliphatic or heterocyclic moieties, may optionally be substituted with substituents including, but not limited to aliphatic; heteroaliphatic; aryl; heteroaryl; alkylaryl; alkylheteroaryl; alkoxy; aryloxy; heteroalkoxy; heteroaryloxy; alkylthio; arylthio; heteroalkylthio; heteroarylthio; F; Cl; Br; I; -OH; -NO₂; -CN; -CF₃; -CH₂CF₃; -CHC1₂; -CH₂OH; -CH₂CH₂OH; -CH₂NH₂; -CH₂SO₂CH₃; -C(O)R_x; -CO₂(R_x); -CON(R_x)₂; -OC(O)R_x; -OCO₂R_x; -OCON(R_x)₂; -N(R_x)₂; -S(O)₂R_x; -NR_x(CO)R_x wherein each occurrence of R_x independently includes, but is not limited to, aliphatic, heteroaliphatic, aryl, heteroaryl, alkylaryl, or alkylheteroaryl, wherein any of the aliphatic, heteroaliphatic, alkylaryl, or alkylheteroaryl substituents described above and herein may be substituted or unsubstituted, branched or unbranched, cyclic or acyclic, and wherein any of the aryl or heteroaryl substituents described above and herein may be substituted or unsubstituted. Additional examples of generally

applicable substituents are illustrated by the specific embodiments shown in the Examples that are described herein.

[0024] The term "heteroaliphatic", as used herein, refers to aliphatic moieties in which one or more carbon atoms in the main chain have been substituted with a heteroatom. Thus, a heteroaliphatic group refers to an aliphatic chain which contains one or more oxygen, sulfur, nitrogen, phosphorus or silicon atoms, e.g., in place of carbon atoms. Heteroaliphatic moieties may be branched or linear unbranched. In certain embodiments, heteroaliphatic moieties are substituted by independent replacement of one or more of the hydrogen atoms thereon with one or more moieties including, but not limited to aliphatic; alicyclic; heteroaliphatic; heteroalicyclic; aryl; heteroaryl; alkylaryl; alkylheteroaryl; alkoxy; aryloxy; heteroalkoxy; heteroaryloxy; alkylthio; arylthio; heteroalkylthio; heteroarylthio; F; Cl; Br; I; -OH; -NO₂; -CN; -CF₃; -CH₂CF₃; -CHCl₂; -CH₂OH; -CH₂CH₂OH; -CH₂NH₂; -CH₂SO₂CH₃; -C(O)R_x; -CO₂(R_x); -CON(R_x)₂; -OC(O)R_x; -OCO₂R_x; -OCON(R_x)₂; -N(R_x)₂; -S(O)₂R_x; -NR_x(CO)R_x wherein each occurrence of R_x independently includes, but is not limited to, aliphatic, alicyclic, heteroaliphatic, heteroalicyclic, aryl, heteroaryl, alkylaryl, or alkylheteroaryl, wherein any of the aliphatic, alicyclic, heteroaliphatic, heteroalicyclic, alkylaryl, or alkylheteroaryl substituents described above and herein may be substituted or unsubstituted, branched or unbranched, cyclic or acyclic, and wherein any of the aryl or heteroaryl substituents described above and herein may be substituted or unsubstituted. Additional examples of generally applicable substituents are illustrated by the specific embodiments shown in the Examples that are described herein.

[0025] The term "heteroalicyclic", as used herein, refers to compounds which combine the properties of heteroaliphatic and cyclic compounds and include but are not limited to saturated and unsaturated mono- or polycyclic heterocycles such as morpholino, pyrrolidinyl, furanyl, thifuranyl, pyrrolyl etc., which are optionally substituted with one or more functional groups, as defined herein.

[0026] Additionally, it will be appreciated that any of the alicyclic or heteroalicyclic moieties described above and herein may comprise an aryl or heteroaryl moiety fused thereto. Additional examples of generally applicable substituents are illustrated by the specific embodiments shown in the Examples that are described herein.

[0027] The terms "halo" and "halogen" as used herein refer to an atom selected from fluorine, chlorine, bromine and iodine.

[0028] The term "haloalkyl" denotes an alkyl group, as defined above, having one, two, or three halogen atoms attached thereto and is exemplified by such groups as chloromethyl, bromoethyl, trifluoromethyl, and the like.

[0029] The term "heterocycloalkyl" or "heterocycle", as used herein, refers to a non-aromatic 5-, 6- or 7- membered ring or a polycyclic group, including, but not limited to a bi- or tri-cyclic group comprising fused six-membered rings having between one and three heteroatoms independently selected from oxygen, sulfur and nitrogen, wherein (i) each 5-membered ring has 0 to 1 double bonds and each 6-membered ring has 0 to 2 double bonds, (ii) the nitrogen and sulfur heteroatoms may be optionally be oxidized, (iii) the nitrogen heteroatom may optionally be quaternized, and (iv) any of the above heterocyclic rings may be fused to an aryl or heteroaryl ring. Representative heterocycles include, but are not limited to, pyrrolidinyl, pyrazolinyl, pyrazolidinyl, imidazolinyl, imidazolidinyl, piperidinyl, piperazinyl, oxazolidinyl, isoxazolidinyl, morpholinyl, thiazolidinyl, isothiazolidinyl, and tetrahydrofuryl. In certain embodiments, a "substituted heterocycloalkyl or heterocycle" group is utilized and as used herein, refers to a heterocycloalkyl or heterocycle group, as defined above, substituted by the independent replacement of one, two or three of the hydrogen atoms thereon with but are not limited to aliphatic; heteroaliphatic; aryl; heteroaryl; alkylaryl; alkylheteroaryl; alkoxy; aryloxy; heteroalkoxy; heteroaryloxy; alkylthio; arylthio; heteroalkylthio; heteroarylthio; F; Cl; Br; I; -OH; -NO₂; -CN; -CF₃; -CH₂CF₃; -CHCl₂; -CH₂OH; -CH₂CH₂OH; -CH₂NH₂; -CH₂SO₂CH₃; -C(O)R_x; -CO₂(R_x); -CON(R_x)₂; -OC(O)R_x; -OCO₂R_x; -OCON(R_x)₂; -N(R_x)₂; -S(O)₂R_x; -NR_x(CO)R_x wherein each occurrence of R_x independently includes, but is not limited to, aliphatic, heteroaliphatic, aryl, heteroaryl, alkylaryl, or alkylheteroaryl, wherein any of the aliphatic, heteroaliphatic, alkylaryl, or alkylheteroaryl substituents described above and herein may be substituted or unsubstituted, branched or unbranched, cyclic or acyclic, and wherein any of the aryl or heteroaryl substituents described above and herein may be substituted or unsubstituted. Additional examples or generally applicable substituents are illustrated by the specific embodiments shown in the Examples, which are described herein.

[0030] As used herein, the terms "aliphatic", "heteroaliphatic", "alkyl", "alkenyl", "alkynyl", "heteroalkyl", "heteroalkenyl", "heteroalkynyl", and the like encompass substituted and unsubstituted, saturated and unsaturated, and linear and branched groups. Similarly, the terms " Alicyclic", "heteroalicyclic", "heterocycloalkyl", "heterocycle" and the like encompass

substituted and unsubstituted, and saturated and unsaturated groups. Additionally, the terms "cycloalkyl", "cycloalkenyl", "cycloalkynyl", "heterocycloalkyl", "heterocycloalkenyl", "heterocycloalkynyl", "aryl", "heteroaryl" and the like encompass both substituted and unsubstituted groups.

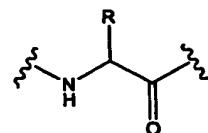
[0031] It will be appreciated that additional examples of generally applicable substitutents are illustrated by the specific embodiments shown in the Examples which are described herein, but are not limited to these Examples.

[0032] By the term "protecting group", has used herein, it is meant that a particular functional moiety, e.g., O, S, or N, is temporarily blocked so that a reaction can be carried out selectively at another reactive site in a multifunctional compound. In preferred embodiments, a protecting group reacts selectively in good yield to give a protected substrate that is stable to the projected reactions; the protecting group must be selectively removed in good yield by readily available, preferably nontoxic reagents that do not attack the other functional groups; the protecting group forms an easily separable derivative (more preferably without the generation of new stereogenic centers); and the protecting group has a minimum of additional functionality to avoid further sites of reaction. As detailed herein, oxygen, sulfur, nitrogen and carbon protecting groups may be utilized. For example, in certain embodiments, as detailed herein, certain exemplary oxygen protecting groups are utilized. These oxygen protecting groups include, but are not limited to methyl ethers, substituted methyl ethers (e.g., MOM (methoxymethyl ether), MTM (methylthiomethyl ether), BOM (benzyloxymethyl ether), PMBM or MPM (p-methoxybenzyloxymethyl ether), to name a few), substituted ethyl ethers, substituted benzyl ethers, silyl ethers (e.g., TMS (trimethylsilyl ether), TES (triethylsilylether), TIPS (triisopropylsilyl ether), TBDMS (t-butyldimethylsilyl ether), tribenzyl silyl ether, TBDPS (t-butyldiphenyl silyl ether), to name a few), esters (e.g., formate, acetate, benzoate (Bz), trifluoroacetate, dichloroacetate, to name a few), carbonates, cyclic acetals and ketals. In certain other exemplary embodiments, nitrogen protecting groups are utilized. These nitrogen protecting groups include, but are not limited to, carbamates (including methyl, ethyl and substituted ethyl carbamates (e.g., Troc), to name a few) amides, cyclic imide derivatives, N-Alkyl and N-Aryl amines, imine derivatives, and enamine derivatives, to name a few. Certain other exemplary protecting groups are detailed herein, however, it will be appreciated that the present invention is not intended to be limited to these protecting groups; rather, a variety of

additional equivalent protecting groups can be readily identified using the above criteria and utilized in the present invention. Additionally, a variety of protecting groups are described in "Protective Groups in Organic Synthesis" Third Ed. Greene, T.W. and Wuts, P.G., Eds., John Wiley & Sons, New York: 1999, the entire contents of which are hereby incorporated by reference.

[0033] The term "natural amino acid side chain" as used herein refers to the side chain of any one of the common, naturally occurring L-amino acids found in naturally occurring proteins: glycine (Gly), alanine (Ala), valine (Val), leucine (Leu), isoleucine (Ile), lysine (Lys), arginine (Arg), histidine (His), proline (Pro), serine (Ser), threonine (Thr), phenylalanine (Phe), tyrosine (Tyr), tryptophan (Trp), aspartic acid (Asp), glutamic acid (Glu), asparagine (Asn), glutamine (Gln), cysteine (Cys) and methionine (Met).

[0034] The term "unnatural amino acid side chain" as used herein refers to the side chain of all amino acids which are not natural amino acids. This includes, for example, α -, β -, D-, L-



amino acid residues, and compounds of the general formula wherein the side chain R is other than the amino acid side chains occurring in nature.

[0035] More generally, the term "amino acid side chain", as used herein, encompasses natural amino acid and unnatural amino acid side chains.

[0036] As used herein, the term "pharmaceutically useful group or entity" refers to a compound or fragment thereof, or an organic moiety which, when covalently attached to a peptide or protein, can exert some biological or diagnostic function or activity when administered to a subject, or enhance the therapeutic, diagnostic or preventive properties of the parent peptide and/or protein in biomedical applications, or improve safety, alter biodegradation or excretion, or is detectable. Examples of suitable pharmaceutically useful groups or entities include hydrophilicity/hydrophobicity modifiers, pharmacokinetic modifiers, biologically active modifiers, detectable modifiers. A modifier can have one or more pharmaceutical functions, e.g., biological activity and/or pharmacokinetics modification. Pharmacokinetics modifiers can include, for example, antibodies, antigens, receptor ligands, hydrophilic, hydrophobic or charged groups. Biologically active modifiers include, for example, therapeutic drugs and prodrugs, antigens, immunomodulators. Detectable modifiers include diagnostic labels, such as

radioactive, fluorescent, paramagnetic, superparamagnetic, ferromagnetic, X-ray modulating, X-ray-opaque, ultrasound-reflective, and other substances detectable by one of available clinical or laboratory methods, e.g., scintigraphy, NMR spectroscopy, MRI, X-ray tomography, sonotomography, photoimaging, radioimmunoassay. Modifiers can be small molecules or macromolecules, and can belong to any chemical or pharmaceutical class, e.g., nucleotides, chemotherapeutic agents, antibacterial agents, antiviral agents, immunomodulators, hormones or analogs thereof, enzymes, inhibitors, alkaloids and therapeutic radionuclides. Viral and non-viral gene vectors are considered to be a pharmaceutically useful entity or group.

[0037] The term "biomolecules", as used herein, refers to molecules (e.g., proteins, amino acids, peptides, polynucleotides, nucleotides, carbohydrates, sugars, lipids, nucleoproteins, glycoproteins, lipoproteins, steroids, etc.) which belong to classes of chemical compounds, whether naturally-occurring or artificially created (e.g., by synthetic or recombinant methods), that are commonly found in cells and tissues. Specific classes of biomolecules include, but are not limited to, enzymes, receptors, neurotransmitters, hormones, cytokines, cell response modifiers such as growth factors and chemotactic factors, antibodies, vaccines, haptens, toxins, interferons, ribozymes, anti-sense agents, plasmids, DNA, and RNA.

[0038] As used herein, the term "small molecule" refers to molecules, whether naturally-occurring or artificially created (e.g., via chemical synthesis) that have a relatively low molecular weight. Preferred small molecules are biologically active in that they produce a local or systemic effect in animals, preferably mammals, more preferably humans. Typically, small molecules have a molecular weight of less than about 1500 g/mol. In certain preferred embodiments, the small molecule is a drug. Preferably, though not necessarily, the drug is one that has already been deemed safe and effective for use by the appropriate governmental agency or body. For example, drugs for human use listed by the FDA under 21 C.F.R. §§ 330.5, 331 through 361, and 440 through 460; drugs for veterinary use listed by the FDA under 21 C.F.R. §§ 500 through 589, incorporated herein by reference, are all considered suitable for use with the present hydrophilic polymers.

[0039] Classes of small molecule drugs that can be used in the practice of the present invention include, but are not limited to, vitamins, anti-AIDS substances, anti-cancer substances, antibiotics, immunosuppressants, anti-viral substances, enzyme inhibitors, neurotoxins, opioids, hypnotics, anti-histamines, lubricants, tranquilizers, anti-convulsants, muscle relaxants and

anti-Parkinson substances, anti-spasmodics and muscle contractants including channel blockers, miotics and anti-cholinergics, anti-glaucoma compounds, anti-parasite and/or anti-protozoal compounds, modulators of cell-extracellular matrix interactions including cell growth inhibitors and anti-adhesion molecules, vasodilating agents, inhibitors of DNA, RNA or protein synthesis, anti-hypertensives, analgesics, anti-pyretics, steroid and non-steroidal anti-inflammatory agents, anti-angiogenic factors, anti-secretory factors, anticoagulants and/or antithrombotic agents, local anesthetics, ophthalmics, prostaglandins, anti-depressants, anti-psychotic substances, anti-emetics, imaging agents. Many large molecules are also drugs.

[0040] A more complete, although not exhaustive, listing of classes and specific drugs suitable for practicing the present invention may be found in "Pharmaceutical Substances: Syntheses, Patents, Applications" by Axel Kleemann and Jurgen Engel, Thieme Medical Publishing, 1999 and the "Merck Index: An Encyclopedia of Chemicals, Drugs, and Biologicals", Edited by Susan Budavari *et al.*, CRC Press, 1996, both of which are incorporated herein by reference.

[0041] As used herein, the term "macromolecules" refers to molecules, whether naturally-occurring or artificially created (*e.g.*, via chemical synthesis) that have a relatively high molecular weight, *e.g.*, generally above 1500 g/mole. Preferred macromolecules are biologically active in that they exert a biological function in animals, preferably mammals, more preferably humans. Examples of macromolecules include proteins, enzymes, growth factors, cytokines, peptides, polypeptides, polylysine, proteins, lipids, polyelectrolytes, immunoglobulins, DNA, RNA, ribozymes, plasmids, and lectins. For the purpose of this invention, supramolecular constructs such as viruses and protein associates (*e.g.*, dimers) are considered to be macromolecules. When covalently attached to a peptide or protein, a macromolecule may be chemically modified prior to being covalently attached to said peptide or protein.

[0042] As used herein, the term "diagnostic label" refers to an atom, group of atoms, moiety or functional group, a nanocrystal, or other discrete element of a composition of matter, that can be detected *in vivo* or *ex vivo* using analytical methods known in the art. When covalently attached to a peptide or protein, such diagnostic labels permit the monitoring of the peptide or protein *in vivo*. On the other hand, constructs and compositions that include diagnostic labels can be used to monitor biological functions or structures. Examples of diagnostic labels include, without limitations, labels that can be used in medical diagnostic procedures, such as,

radiopharmaceutical or radioactive isotopes for gamma scintigraphy and Positron Emission Tomography (PET), contrast agent for Magnetic Resonance Imaging (MRI) (for example paramagnetic atoms and superparamagnetic nanocrystals), contrast agent for computed tomography, contrast agent for X-ray imaging method, agent for ultrasound diagnostic method, agent for neutron activation, and moiety which can reflect, scatter or affect X-rays, ultrasounds, radiowaves and microwaves, fluorophores in various optical procedures, etc.

BRIEF DESCRIPTION OF THE DRAWING

[0043] Figure 1 depicts examples of bifunctional glycopeptides (*i.e.*, diglycosylated peptides) containing *O*-linkages prepared by the method of the invention.

[0044] Figure 2 depicts anomeric β -glycosylamines (22 and 23) that differ at a single stereocenter (asterisks), and a bifunctional glycopeptide (24) containing one of these structurally complex glycans.

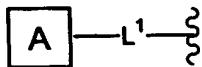
DETAILED DESCRIPTION OF CERTAIN PREFERRED EMBODIMENTS OF THE INVENTION

[0045] In one aspect, the present invention provides novel methodologies for the synthesis of peptides and/or proteins functionalized at two or more non-adjacent amino acid sites (*e.g.*, bearing a pharmaceutically useful group or entity covalently attached to at least two non-adjacent amino acid residues of the polypeptide/protein chain). In certain embodiments, the invention provides a system for the synthesis of polypeptides and proteins bearing at least two glycans covalently attached to non-adjacent amino acid residues of the polypeptide/protein chain. In certain embodiments, in the context of synthetic studies developed for the total synthesis of glycosylated fragments of erythropoietin, generalized methodologies were developed for the improved synthesis of polyglycosylated peptides and proteins. This general synthetic method encompasses the realization that native chemical ligation (NCL) is a glycan-compatible process that may be used for assembling large multiply glycosylated polypeptides from glycopeptide building blocks. In yet another aspect, the present invention also provides the recognition that chemoselective reaction of a glycopeptide containing a C-terminal cysteine with a glycopeptide thioester may be achieved to from the corresponding diglycosylated peptide adduct.

[0046] Specific examples, particularly with respect to the synthesis of bifunctional glycopeptides (*i.e.*, diglycosylated peptides), are described in more detail below and in the Exemplification herein, along with certain general methodologies developed during the course of these syntheses. It will be appreciated by one of ordinary skill in the art that these examples are not intended to be limiting; rather all equivalents are intended to be incorporated into the scope of the present invention. In particular, the inventive method may be generally adapted to the preparation of polyfunctionalized peptides and proteins.

[0047] 1) Inventive method

[0048] In one aspect of the present invention, there is provided a method for preparing a polyfunctionalized peptide comprising a peptidic backbone made up of four or more amino acids wherein two or more non-adjacent amino acids are independently substituted with a moiety having the structure:



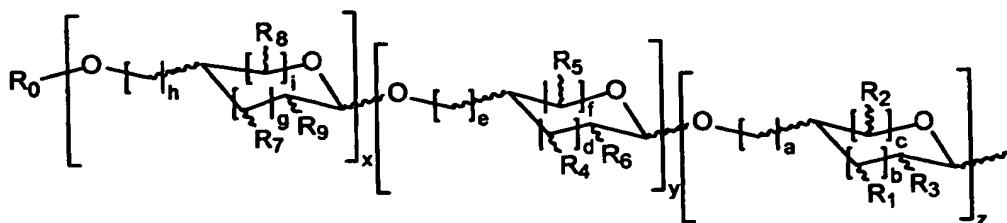
wherein each occurrence of L¹ is independently a substituted or unsubstituted, linear or branched, cyclic or acyclic, saturated or unsaturated aliphatic or heteroaliphatic moiety;

each occurrence of A is independently an aliphatic, heteroaliphatic, aryl, heteroaryl or a pharmaceutically useful group or entity;

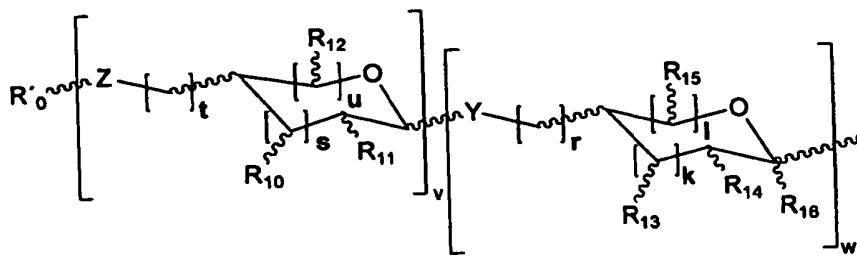
with the proviso that the peptide sequence between any two consecutive, non-adjacent, amino acids bearing a A-L¹-moiety comprises at least one cysteine residue.

[0049] In certain embodiments, each occurrence of A is independently a pharmaceutically useful group or entity. In certain embodiments, each occurrence of A is independently a biomolecule, a small molecule, a macromolecule or a diagnostic label.

[0050] In certain exemplary embodiments, each occurrence of A is independently a carbohydrate determinant having the structure:



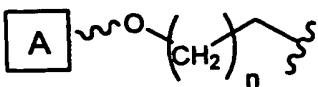
wherein a, b, c, d, e, f, g, h, i, x, y and z are independently 0, 1, 2 or 3, with the proviso that the x, y and z bracketed structures represent furanose or pyranose moieties and the sum of b and c is 1 or 2, the sum of d and f is 1 or 2, and the sum of g and i is 1 or 2, and with the proviso that x, y and z are not simultaneously 0; wherein R₀ is hydrogen, a linear or branched chain alkyl, acyl, arylalkyl or aryl group; wherein each occurrence of R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈ and R₉ is independently hydrogen, OH, ORⁱ, NHRⁱ, NHCORⁱ, F, CH₂OH, CH₂ORⁱ, a substituted or unsubstituted linear or branched chain alkyl, (mono-, di- or tri)hydroxyalkyl, (mono-, di- or tri)acyloxyalkyl, arylalkyl or aryl group; wherein each occurrence of Rⁱ is independently hydrogen, CHO, COORⁱⁱ, or a substituted or unsubstituted linear or branched chain alkyl, acyl, arylalkyl or aryl group or a saccharide moiety having the structure:



wherein Y and Z are independently NH or O; wherein k, l, r, s, t, u, v and w are each independently 0, 1 or 2; with the proviso that the v and w bracketed structures represent furanose or pyranose moieties and the sum of l and k is 1 or 2, and the sum of s and u is 1 or 2, and with the proviso that v and w are not simultaneously 0; wherein R'0 is hydrogen, a linear or branched chain alkyl, acyl, arylalkyl or aryl group; wherein each occurrence of R₁₀, R₁₁, R₁₂, R₁₃, R₁₄ and R₁₅ is independently hydrogen, OH, ORⁱⁱⁱ, NHRⁱⁱⁱ, NHCORⁱⁱⁱ, F, CH₂OH, CH₂ORⁱⁱⁱ, or a substituted or unsubstituted linear or branched chain alkyl, (mono-, di- or tri)hydroxyalkyl, (mono-, di- or tri)acyloxyalkyl, arylalkyl or aryl group; wherein each occurrence of R₁₆ is hydrogen, COOH, COORⁱⁱ, CONHRⁱⁱ, a substituted or unsubstituted linear or branched chain alkyl or aryl group; wherein each occurrence of Rⁱⁱⁱ is hydrogen, CHO, COOR^{iv}, or a substituted or unsubstituted linear or branched chain alkyl, acyl, arylalkyl or aryl group; and wherein each occurrence of Rⁱⁱ and R^{iv} are each independently H, or a substituted or unsubstituted linear or branched chain alkyl, arylalkyl or aryl group.

[0051] In certain embodiments, each occurrence of L¹ is independently -O-(CH₂)_n- , wherein n is 0-9, or a glycoside-containing moiety (e.g mono- or poly-saccharide).

[0052] In certain embodiments, L¹ is -O-(CH₂)_n-CH₂- and two or more non-adjacent amino acids is/are independently substituted with a moiety having the structure:



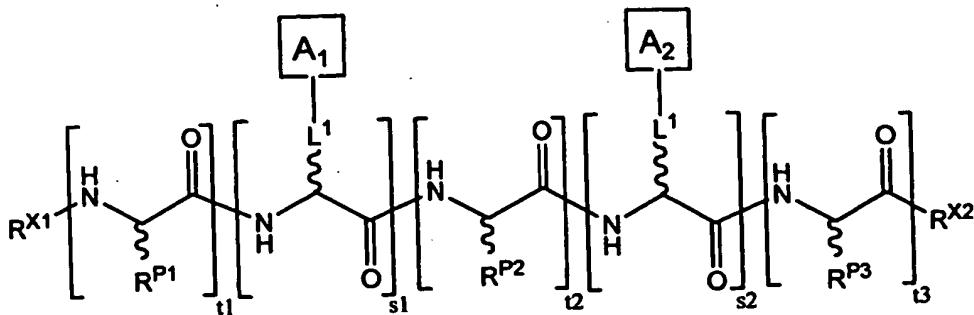
wherein each occurrence of n is independently 0-8; and A is as defined above. In certain other embodiments, A is a carbohydrate moiety and each occurrence of the n-alkyl glycosidic moiety is independently either α- or β-linked to an amino acid residue of the backbone. It will be appreciated that polyfunctionalized peptides made according to the method of the present invention are not limited to those where each occurrence of L¹ comprises n-alkyl where n is greater than or equal to 1; rather each occurrence of A can be independently linked via the traditional direct linkage (n=0), via n-alkyl (such as pentyl), via a mono-saccharide moiety or any combination thereof. In preferred embodiments, each occurrence of A is independently selected from the group consisting of Globo-H, fucosyl GM1, KH-1, glycophorin, STN, (2,3)ST, Le^y, Le^x, N3, Tn, 2,6-STn, Gb3 and TF.

[0053] In certain embodiments, occurrences of A may be the same or different.

[0054] In certain other embodiments, certain occurrences of A are clustered. For example, in certain embodiments, the polyfunctionalized peptide comprises at least two adjacent functionalized amino acids (*i.e.*, clustered functionalized amino acids), separated from another functionalized site by a peptide amino acid sequence comprising at least one cysteine residue.

[0055] In certain other embodiments, certain occurrences of A are clustered glycosides. For example, in certain embodiments, the multglycosylated peptide comprises at least two adjacent glycosylated amino acids (*i.e.*, clustered glycosylated amino acids), separated from another glycosylated site by a peptide amino acid sequence comprising at least one cysteine residue.

[0056] In certain embodiments, the present invention provides a method for preparing a polyfunctionalized peptide having the structure:



s₁ and s₂ are independently an integer from 1 to about 20;

t₁, t₂ and t₃ are each independently an integer;

R^{X1} is hydrogen, alkyl, acyl, aryl, heteroaryl, -alkyl(aryl), -alkyl(heteroaryl), a nitrogen protecting group, an amino acid or a protected amino acid;

R^{X2} is -OR^{X2a} or -NR^{X2b}R^{X2c}, wherein R^{X2a} is hydrogen, alkyl, aryl, heteroaryl, -alkyl(aryl), -alkyl(heteroaryl), a carboxylic acid protecting group, an amino acid or a protected amino acid; and R^{X2b} and R^{X2c} are independently hydrogen, alkyl, aryl, heteroaryl, -alkyl(aryl), -alkyl(heteroaryl), a nitrogen protecting group, an amino acid or a protected amino acid;

R^{P1}, R^{P2} and R^{P3} are independently H, alkyl, heteroalkyl, aryl, heteroaryl, -alkyl(aryl), -alkyl(heteroaryl), or a natural or non-natural amino acid side chain;

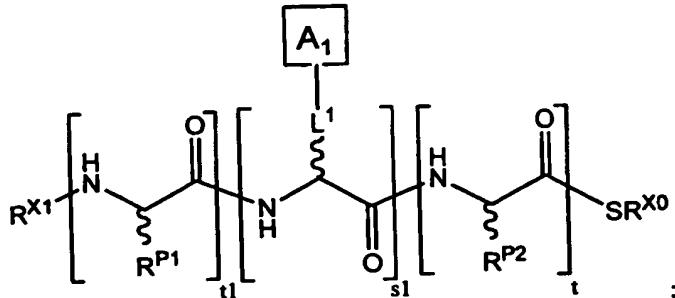
each occurrence of L¹ is independently a substituted or unsubstituted aliphatic or heteroaliphatic moiety;

A₁ and A₂ are each independently an aliphatic, heteroaliphatic, aryl, heteroaryl or a pharmaceutically useful group or entity; and

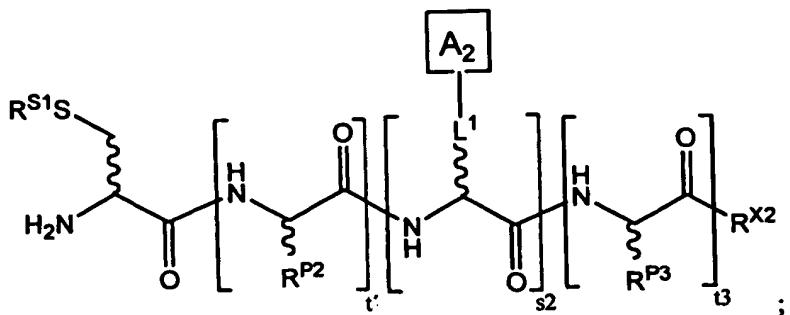
at least one occurrence of the bracketed structure t₂ is a cysteine residue or protected cysteine residue;

wherein the method comprises a step of:

reacting a peptide acyl donor having the structure:



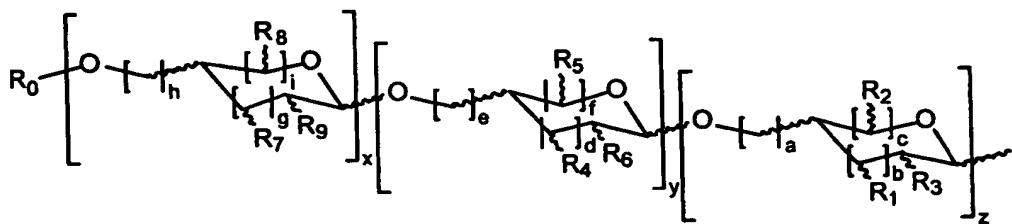
with a peptide amine acceptor having the structure:



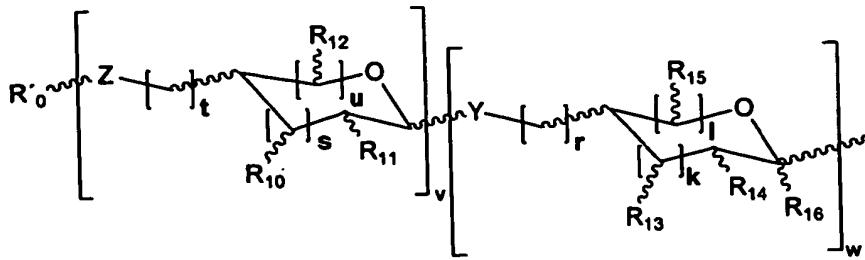
under suitable conditions to effect disulfide exchange ligation;

wherein the sum $t+t'$ equals $(t_2)+1$; R^{S1} is a sulfide protecting group; and R^{X0} is a group such that the moiety $-C(=O)SR^{X0}$ can be made to undergo disulfide exchange with the glycopeptide amine acceptor.

In certain exemplary embodiments, A_1 and A_2 are each independently a carbohydrate domain having the structure:

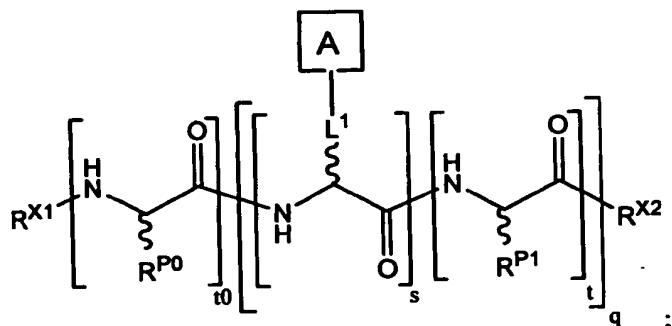


wherein a , b , c , d , e , f , g , h , i , x , y and z are independently 0, 1, 2 or 3, with the proviso that the x , y and z bracketed structures represent furanose or pyranose moieties and the sum of b and c is 1 or 2, the sum of d and f is 1 or 2, and the sum of g and i is 1 or 2, and with the proviso that x , y and z are not simultaneously 0; wherein R_0 is hydrogen, a linear or branched chain alkyl, acyl, arylalkyl or aryl group; wherein each occurrence of R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 and R_9 is independently hydrogen, OH, OR^i , NHR^i , $NHCOR^i$, F, CH_2OH , CH_2OR^i , a substituted or unsubstituted linear or branched chain alkyl, (mono-, di- or tri)hydroxyalkyl, (mono-, di- or tri)acyloxyalkyl, arylalkyl or aryl group; wherein each occurrence of R^i is independently hydrogen, CHO, $COOR^{ii}$, or a substituted or unsubstituted linear or branched chain alkyl, acyl, arylalkyl or aryl group or a saccharide moiety having the structure:



[0057] wherein Y and Z are independently NH or O; wherein k, l, r, s, t, u, v and w are each independently 0, 1 or 2; with the proviso that the v and w bracketed structures represent furanose or pyranose moieties and the sum of l and k is 1 or 2, and the sum of s and u is 1 or 2, and with the proviso that v and w are not simultaneously 0; wherein R'0 is hydrogen, a linear or branched chain alkyl, acyl, arylalkyl or aryl group; wherein each occurrence of R₁₀, R₁₁, R₁₂, R₁₃, R₁₄ and R₁₅ is independently hydrogen, OH, ORⁱⁱⁱ, NHRⁱⁱⁱ, NHCORⁱⁱⁱ, F, CH₂OH, CH₂ORⁱⁱⁱ, or a substituted or unsubstituted linear or branched chain alkyl, (mono-, di- or tri)hydroxyalkyl, (mono-, di- or tri)acyloxyalkyl, arylalkyl or aryl group; wherein each occurrence of R₁₆ is hydrogen, COOH, COORⁱⁱ, CONHRⁱⁱ, a substituted or unsubstituted linear or branched chain alkyl or aryl group; wherein each occurrence of Rⁱⁱⁱ is hydrogen, CHO, COOR^{iv}, or a substituted or unsubstituted linear or branched chain alkyl, acyl, arylalkyl or aryl group; and wherein each occurrence of Rⁱⁱ and R^{iv} are each independently H, or a substituted or unsubstituted linear or branched chain alkyl, arylalkyl or aryl group; and wherein each glycosidic moiety is either α - or β -linked to an amino acid.

[0058] In certain embodiments, the step may be carried out once, or repeated a desired number of times, to prepare a polyfunctionalized peptide having the structure:



wherein R^{X1} and R^{X2} are as defined above;

each occurrence of A may be the same or different and may be as defined for A₁ and A₂ above;

each occurrence of R^{P1} may be the same or different and may be as defined for R^{P1} and R^{P2} above;

q is an integer greater than or equal to 2;

each occurrence of s is independently an integer from 1 to about 20;

each occurrence of t is independently an integer;

t₀ is an integer; and

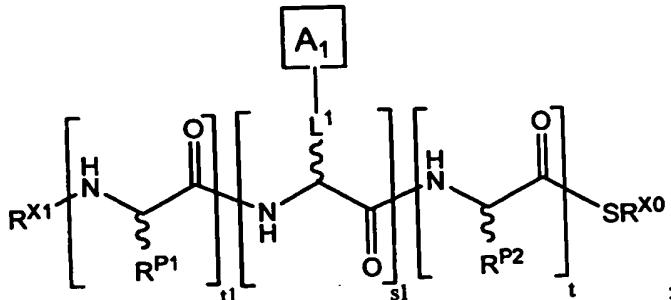
each occurrence of R^{P0} is independently H, alkyl, heteroalkyl, aryl, heteroaryl, -alkyl(aryl), -alkyl(heteroaryl), or a natural or non-natural amino acid side chain.

[0059] In certain embodiments, q is an integer between 2 and about 5. In certain embodiments, q is an integer between 2 and about 10. In certain embodiments, q is an integer between 2 and about 15. In certain embodiments, q is an integer between 2 and about 20. In certain embodiments, q is an integer between 2 and about 25. In certain embodiments, q is an integer between 2 and about 30. In certain embodiments, q is an integer greater than 30. In certain embodiments, q is 2.

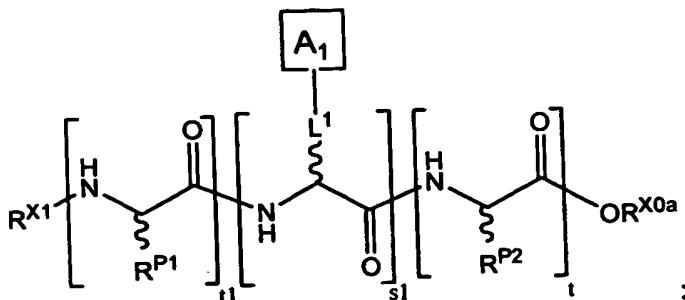
[0060] In certain embodiments, the sum s+t is between about 2 and about 6. In certain embodiments, the sum s+t is between about 2 and about 10. In certain embodiments, the sum s+t is between about 2 and about 15. In certain embodiments, the sum s+t is between about 2 and about 20. In certain embodiments, the sum s+t is between about 2 and about 50. In certain embodiments, the sum s+t is between about 2 and about 100. In certain embodiments, the sum s+t is between about 2 and about 150. In certain embodiments, the sum s+t is between about 2 and about 200. In certain embodiments, the sum s+t may be greater than 200.

[0061] In certain embodiments, t₀ is an integer from 0 to about 2. In certain embodiments, t₀ is an integer from 0 to about 5. In certain embodiments, t₀ is an integer from 0 to about 10. In certain embodiments, t₀ is an integer from 0 to about 15. In certain embodiments, t₀ is an integer from 0 to about 20. In certain embodiments, t₀ is an integer from 0 to about 25. In certain embodiments, t₀ is an integer from 0 to about 30. In certain embodiments, t₀ is an integer from 0 to about 50. In certain embodiments, t₀ is an integer from 0 to about 100. In certain embodiments, t₀ is an integer from 0 to about 150. In certain embodiments, t₀ is an integer from 0 to about 200. In certain embodiments, t₀ is an integer greater than 200.

- [0062] In certain other embodiments, R^{X1} is hydrogen, Fmoc or Ac.
- [0063] In certain other embodiments, R^{X2} is NH_2 .
- [0064] In certain other embodiments, R^{X0} is $-CH_2CH_2SO_3Na$.
- [0065] In certain other embodiments, R^{S1} is $-StBu$.
- [0066] In certain exemplary embodiments, the peptide acyl donor having the structure:

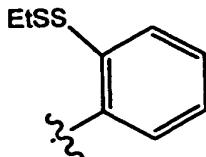


is prepared by reacting $R^{X0}S-Na$ with a compound having the structure:

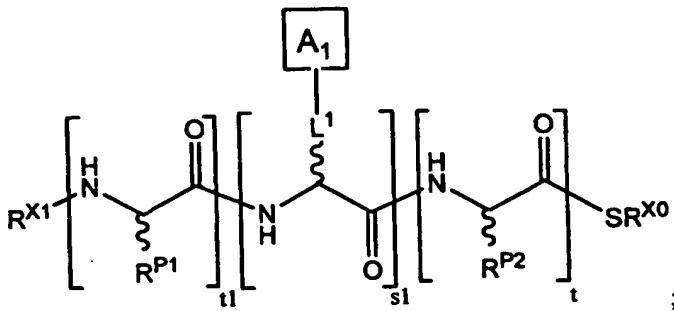


wherein R^{X0a} is a sulfur-substituted aryl moiety.

- [0067] In certain exemplary embodiments, R^{X0a} has the following structure:



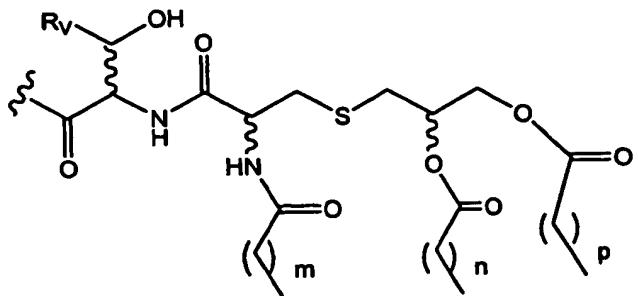
- [0068] In certain exemplary embodiments, in the peptide acyl donor having the structure:



the amino acyl residue directly attached to $-SR^{X0}$ is phenylalanine.

[0069] In certain embodiments, when at least one occurrence of A (or A₁ and/or A₂, as further defined for A) is a carbohydrate domain, some or all of carbohydrate domains are O-linked to the peptide backbone. In certain other embodiments, when at least one occurrence of A (or A₁ and/or A₂, as further defined for A) is a carbohydrate domain, some or all of carbohydrate domains are N-linked to the peptide backbone. In certain embodiments, the peptide sequence between each point of attachment of the A moieties comprises a cysteine residue. In certain embodiments, the polyfunctionalized construct (*i.e.*, construct bearing more than one A moiety) is prepared by Native Chemical Ligation. In certain embodiments, the polyfunctionalized peptides obtained by the inventive method are symmetrical (functionalized peptide building blocks to be ligated have the same peptide sequence (safe for N- and/or C-terminals) and bear the same functionalization pattern), nonsymmetrical (functionalized peptide building blocks to be ligated differ in peptide sequence (excluding N- and C-terminals) and/or functionalization pattern). In certain embodiments, the inventive method allows the preparation of multglycosylated peptides designed to approximate the spatial position(s) of carbohydrate(s) in glycoprotein/glycopeptides of interest (*e.g.*, naturally occurring glycoproteins such as gp120 and erythropoietin).

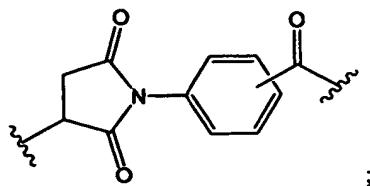
[0070] In certain embodiments, the method further comprises a step of conjugating the polyfunctionalized peptide to an immunogenic carrier. In certain exemplary embodiments, the carrier is a protein, a peptide or a lipid. In certain other exemplary embodiments, the carrier is Bovine Serum Albumin (BSA), Keyhole Limpet Hemocyanin (KLH) or polylysine. In certain other embodiments, the carrier is a lipid carrier having the structure:



wherein m, n and p are each independently integers between about 8 and 20; and Rv is hydrogen, substituted or unsubstituted linear or branched chain lower alkyl or substituted or unsubstituted phenyl. In certain exemplary embodiments, m', n' and p' are each 14 and the lipid is tripalmitoyl-S-glycercylcysteinylserine (e.g., PamCys).

[0071] It will be appreciated that the carrier can be linked to the polyfunctionalized peptide either directly or through a crosslinker, and thus the peptide may be attached to a protein, peptids, or lipid, as well as a (crosslinker-protein), (crosslinker-peptide) and (crosslinker-lipid) moiety.

[0072] Crosslinkers suited to the invention are widely known in the art (see, for example, 1994 Pierce Technical Handbook: cross-linking (Appendix A), which is available at <http://www.piercenet.com/resources/browse.cfm?fldID=184>), including bromoacetic NHS ester, 6-(iodoacetamido)caproic acid NHS ester, maleimidoacetic acid NHS ester, maleimidobenzoic acide NHS ester, etc. In certain preferred embodiments, the crosslinker is MMCCH (4-(maleimidomethyl) cyclohexane-1-carboxyl hydrazide). In certain other preferred embodiments, the crosslinker is MBS (m-maleimidobenzoyl acid N-Hydroxysuccinimidy ester). In certain embodiments, the crosslinker is a fragment having the structure:



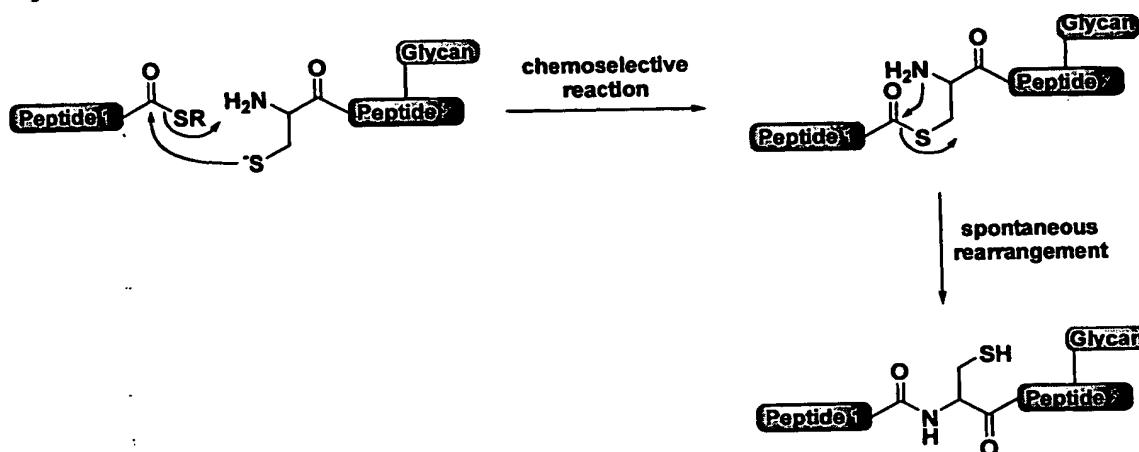
whereby said structure is generated upon conjugation of a maleimidobenzoic acid N-hydroxy succinimide ester with a suitable functionality on the polyfunctionalized peptide.

[0073] 2) Synthetic Methodology

[0074] Native Chemical Ligation

[0075] One of the more widely used methods for the synthesis of glycopeptides is native chemical ligation (NCL).⁴⁸ First reported by Kent in 1994, NCL allows for the assembly of large proteins with native amide bonds from unprotected peptide building blocks (Scheme 1). Furthermore, the reaction is mild, selective, and compatible with the presence of glycans. When glycans are present in the reaction, they are typically found on the C-terminal side. In the event, a glycopeptide containing a C-terminal cysteine undergoes a chemoselective reaction with a peptide thioester. The resulting peptide thioester then rearranges spontaneously to furnish a native peptide bond, effectively lengthening the peptide backbone of the glycopeptide.

[0076] Scheme 1. Exemplary Native Chemical Ligation methodology



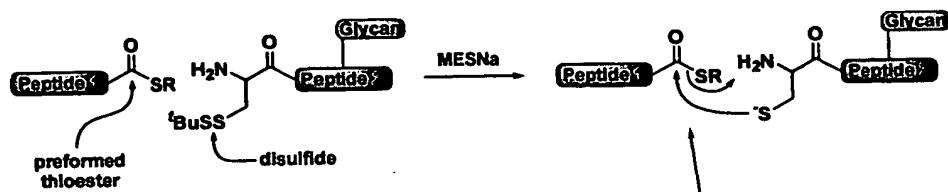
[0077] As discussed above, glycopeptides bearing multiple glycosidic domains are of particular interest. Such structures include the synthesis of gp120 fragments, erythropoietin (EPO), human chorionic gonadotropin (HCG), and amyloid precursor protein (APP). Drawing from previous studies in the area,⁴⁹ NCL was explored as a way to couple two small glycopeptides to form one large glycopeptide. One drawback that became apparent lies in the difficulty with which unprotected glycopeptide α -thioesters are synthesized.⁵⁰ Accordingly, a new method for the synthesis of glycopeptide α -thioesters, or their equivalent, was devised, to allow the assembly of glycopeptides bearing multiple glycosidic domains.

[0078] A closer look at the reaction mechanism of NCL provided clues as to how this problem might be approached. In one embodiment of the reaction itself, the sulphydryl group of the cysteine coupling partner, initially protected as a *t*-butyldisulfide, is released by disulfide exchange with 2-mercaptopethanesulfonic acid (MESNa) and then undergoes trans-

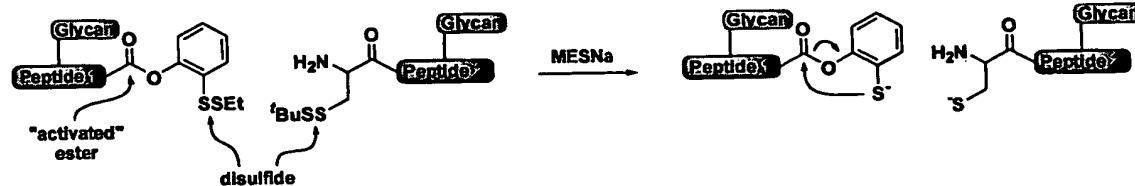
thioesterification as mentioned above. Without wishing to be bound to any particular theory, it was thought that it might be possible to mask the α -thioester as a phenyl ester, bearing an ethyldisulfide in the *ortho* position (Scheme 2) by taking advantage of this disulfide exchange. Under the reaction conditions, the free sulphydryl would be released and undergo a spontaneous rearrangement to furnish a thioester capable of entering into the NCL pathway.

[0079] Scheme 2. A new method for the generation of peptide α -thioesters.

NCL:

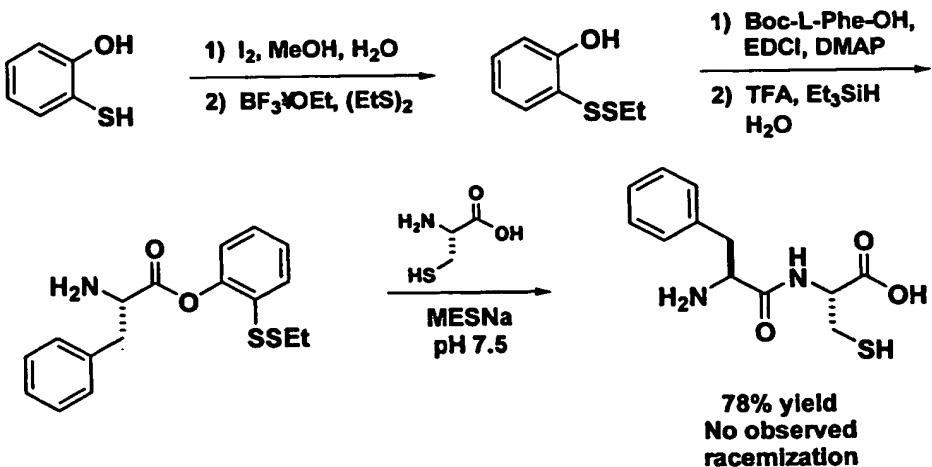


Modification:



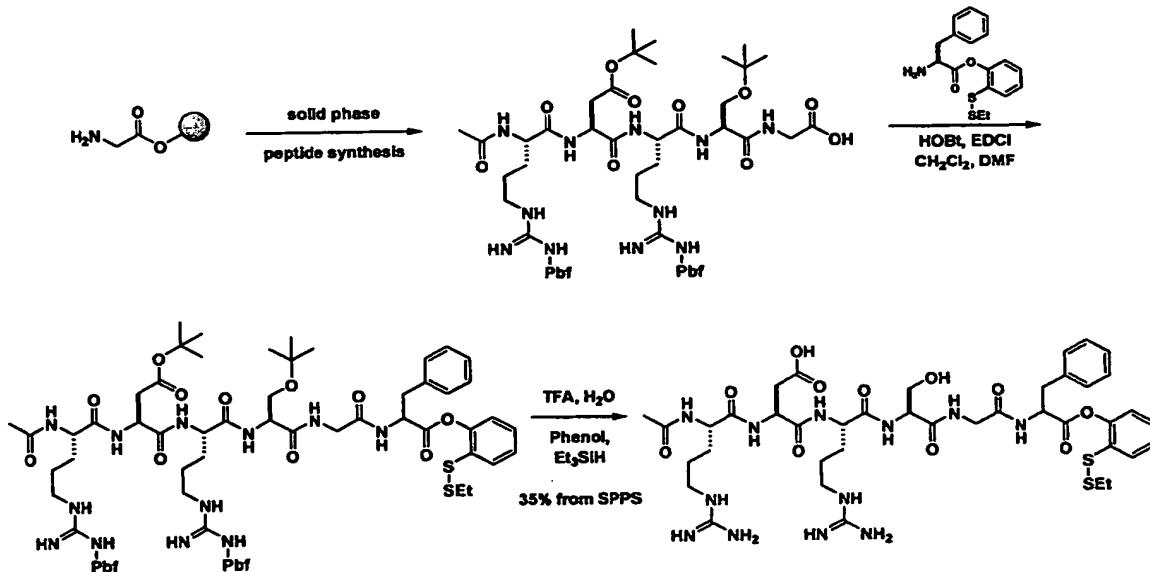
[0080] To test the viability of this proposed modification, the simple dipeptide, PheCys, was synthesized. The desired *ortho*-substituted phenol was synthesized in two steps from commercially available 2-mercaptophenol by oxidation and disulfide exchange. Coupling of the phenol to phenylalanine followed by the removal of the Boc protecting group set the stage for our first attempt (Scheme 3). The reaction proceeded in good yield. More importantly no racemization was observed under the reaction conditions.

[0081] Scheme 3. Exemplary NCL with masked thioester



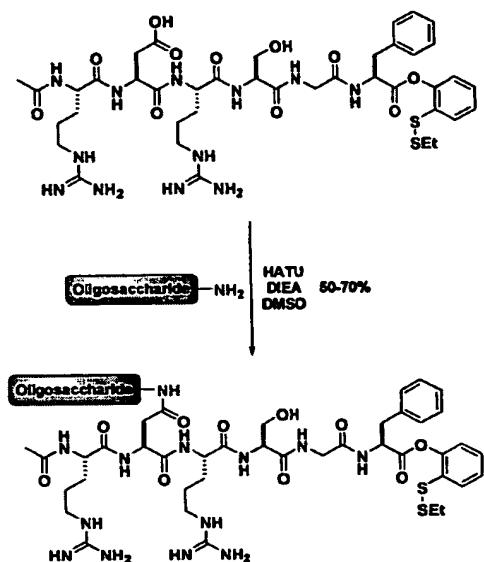
[0082] The synthesis of glycopeptides bearing two glycosidic domains was then undertaken. Synthesis of the *N*-terminal fragment (the fragment that would ultimately become the masked thioester) began with the solid phase synthesis of a peptide acid, followed by standard peptide coupling to introduce the phenylalanine bearing the phenyl ester. TFA deprotection of the side chain protecting groups furnished the peptide backbone for the *N*-terminal fragment (Scheme 4).

[0083] **Scheme 4.** Exemplary synthesis of the peptide backbone of the *N*-terminal fragment.



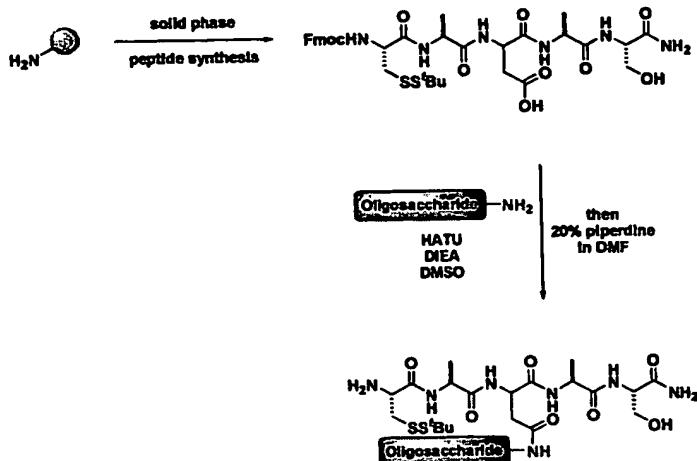
[0084] Coupling of an oligosaccharide with excess peptide furnished a fully deprotected glycopeptide ready for ligation (Scheme 5).

[0085] **Scheme 5.** Exemplary approach to the completion of the *N*-terminal fragment



[0086] With the *N*-terminal portion in hand, the synthesis of the *C*-terminal coupling partner was then carried out. In a similar way, the peptide backbone was prepared by solid phase peptide synthesis. Upon cleavage from the resin, glycosylation and Fmoc removal, the *C*-terminal glycopeptide was obtained (Scheme 6).

[0087] **Scheme 6.** Exemplary synthesis of the *C*-terminal glycopeptide



[0088] In certain exemplary embodiments, the synthesis of glycopeptides bearing two glycosydic domains was accomplished as follows: equal molar amounts of both the *N*-terminal and *C*-terminal glycopeptides were combined in a LCMS vial and to this was added a solution of MESNa in phosphate buffered saline. The reaction was monitored by LCMS and to our amazement, the rearrangement of the phenyl ester to thioester occurred quite rapidly,⁵¹ additionally mass peaks corresponding to the desired product were observed almost instantly. Upon completion of the reaction, any remaining disulfide bonds were reduced by the action of tris(2-carboxyethyl) phosphine hydrochloride (TCEP). The crude reaction mixture was then subjected to HPLC for purification. The reaction proceeded extremely well, providing doubly glycosylated glycopeptides, typically in 60-70% yield. Exemplary di-glycosylated peptides that may be prepared by the method of the present invention are detailed in the Exemplification below. It should be noted that symmetrical, nonsymmetrical and mixed (*N*-linked and *O*-linked) glycopeptides can be used.

[0089] Accordingly, there is provided herein a method for preparing glycopeptides comprising at least two carbohydrate domains covalently attached thereto. In certain embodiments, some or all of carbohydrate domains are *O*-linked. In certain other embodiments, some or all of carbohydrate domains are *N*-linked. In certain embodiments, the glycopeptide comprises two or more carbohydrate domains covalently attached thereto, wherein the glycopeptide sequence between each point of attachment of the carbohydrate domains comprises a cysteine residue. In certain embodiments, the multi-glycan glycopeptide is prepared by Native Chemical Ligation. In certain embodiments, the method allows for coupling where each

coupling partner is a glycopeptide itself. Symmetrical, nonsymmetrical and mixed (N-linked and O-linked) glycopeptides can be obtained. In certain embodiments, the method involves the *in situ* generation of a thioester that is then used immediately in native chemical ligation.

[0090] *Peptide Thioester Synthesis*

[0091] Several methods have been developed for peptide thioester synthesis, including the original "Boc chemistry" (Boc = *tert*-butyloxycarbonyl) method (See, for example, (1) Canne, L. E.; Walker, S. M.; Kent, S. B. H. "A General Method for the Synthesis of Thioester Resin Linkers for Use in the Solid-Phase Synthesis of Peptide Alpha-Thioacids." *Tetrahedron Lett.* 1995, 36, 1217-1220; and (2) Hojo, H.; Aimoto, S. "Polypeptide Synthesis Using the S-Alkyl Thioester of a Partially Protected Peptide Segment--Synthesis of the DNA- Binding Domain of C-Myb Protein (142-193)-NH₂." *Bull. Chem. Soc. Jpn.* 1991, 64, 111-117) and several Fmoc-compatible systems (See, for example, (1) Shin, Y.; Winans, K. A.; Backes, B. J.; Kent, S. B. H.; Ellman, J. A.; Bertozzi, C. R. "Fmoc-based synthesis of peptide-(alpha)thioesters: Application to the total chemical synthesis of a glycoprotein by native chemical ligation." *J. Am. Chem. Soc.* 1999, 121, 11684-11689; (2) Ingenito, R.; Bianchi, E.; Fattori, D.; Pessi, A. "Solid phase synthesis of peptide C-terminal thioesters by Fmoc/t-Bu chemistry." *J. Am. Chem. Soc.* 1999, 121, 11369-11374; (3) Li, X. Q.; Kawakami, T.; Aimoto, S. "Direct preparation of peptide thioesters using an Fmoc solidphase method." *Tetrahedron Lett.* 1998, 39, 8669-8672; (4) Clippingdale, A. B.; Barrow, C. J.; Wade, J. D. "Peptide thioester preparation by Fmoc solid phase peptide synthesis for use in native chemical ligation." *J. Pept. Sci.* 2000, 6, 225-234; and (5) Bu, X. Z.; Xie, G. Y.; Law, C. W.; Guo, Z. H. "An improved deblocking agent for direct Fmoc solidphase synthesis of peptide thioesters." *Tetrahedron Lett.* 2002, 43, 2419-2422). In certain embodiments, the model thioester is a C-terminal glycine thioester, which is locally achiral and cannot be epimerized, and is therefore easy to synthesize. Though the desired GP120 thioester contains an epimerization-prone C-terminal histidine (His) residue, such thioesters have been synthesized previously and have in fact been shown to modulate favorably the rate of NCL (See, for example, Hackeng, T. M.; Griffin, J. H.; Dawson, P. E. "Protein synthesis by native chemical ligation: Expanded scope by using straightforward methodology." *Proc. Natl. Acad. Sci. U. S. A.* 1999, 96, 10068-10073).

[0092] It is understood that, although the discussion above regarding Native Chemical Ligation focuses on the preparation of multglycosylated peptides, the method may readily be

adapted to other polyfunctionalized peptides and/or proteins. For example, peptides and/or proteins functionalized at more than one amino acid site with a pharmaceutically useful group or entity may be prepared by the method of the invention.

EQUIVALENTS

[0093] The representative examples which follow are intended to help illustrate the invention, and are not intended to, nor should they be construed to, limit the scope of the invention. Indeed, various modifications of the invention and many further embodiments thereof, in addition to those shown and described herein, will become apparent to those skilled in the art from the full contents of this document, including the examples which follow and the references to the scientific and patent literature cited herein. In but one illustrative example, protecting groups play an important role in the synthesis of the carbohydrate domains and synthetic conjugates, as described herein; however it will be appreciated by one of ordinary skill in the art that the present invention encompasses the use of various alternate protecting groups known in the art. Those protecting groups used in the disclosure including the Examples below are merely illustrative.

[0094] It should further be appreciated that, unless otherwise indicated, the contents of those cited references are incorporated herein by reference to help illustrate the state of the art. The following examples contain important additional information, exemplification and guidance which can be adapted to the practice of this invention in its various embodiments and the equivalents thereof.

EXEMPLIFICATION

[0095] The method of this invention can be understood further by the examples that illustrate some of the processes by which the inventive method may be practiced. It will be appreciated, however, that these examples do not limit the invention. Variations of the invention, now known or further developed, are considered to fall within the scope of the present invention as described herein and as hereinafter claimed.

[0096] 1) *General Description of Synthetic Methods:*

[0097] The practitioner has a well-established literature of peptide, protein and glycoside chemistry to draw upon, in combination with the information contained herein, for

guidance on synthetic strategies, protecting groups, and other materials and methods useful for practicing the method of this invention.

[0098] The various references cited herein provide helpful background information on preparing complex glycosides, glycosylated peptides and other glycosylated constructs that may be applied and/or adapted to the method of the present invention.

[0099] According to the present invention, any available techniques can be used to practice the method of the invention. For example, a variety of solution phase synthetic methods such as those discussed in detail below may be used. Alternatively or additionally, a variety combinatorial techniques, parallel synthesis and/or solid phase synthetic methods known in the art may be used.

[0100] Starting materials and reagents used in practicing the specific embodiments of the invention detailed below and herein are either available from commercial suppliers such as Aldrich Chemical Company (Milwaukee, WI), Bachem (Torrance, CA), Sigma (St. Louis, MO), or are prepared by methods well known to a person of ordinary skill in the art following procedures described in such references as Fieser and Fieser 1991, "Reagents for Organic Synthesis", vols 1-17, John Wiley and Sons, New York, NY, 1991; Rodd 1989 "Chemistry of Carbon Compounds", vols. 1-5 and supps, Elsevier Science Publishers, 1989; "Organic Reactions", vols 1-40, John Wiley and Sons, New York, NY, 1991; March 2001, "Advanced Organic Chemistry", 5th ed. John Wiley and Sons, New York, NY; and Larock 1990, "Comprehensive Organic Transformations: A Guide to Functional Group Preparations", 2nd ed. VCH Publishers. These schemes are merely illustrative of some embodiments of the present invention, and various modifications to these schemes can be made and will be suggested to a person of ordinary skill in the art having regard to this disclosure.

[0101] In practicing the invention, starting materials, intermediates, and compounds may be isolated and purified using conventional techniques, including filtration, distillation, crystallization, chromatography, and the like. They may be characterized using conventional methods, including physical constants and spectral data.

[0102] Methods for preparing glycopeptides (e.g., O- or N-linked glycopeptides) and for conjugating peptides and glycopeptides to carriers are known in the art. For example, guidance may be found in U.S. Patent No.: 6,660,714; U.S. Patent Application Nos.: 09/641,742; 10/209,618; and 10/728,041; U.S. Provisional Patent Application Nos.: 60/500,161 and

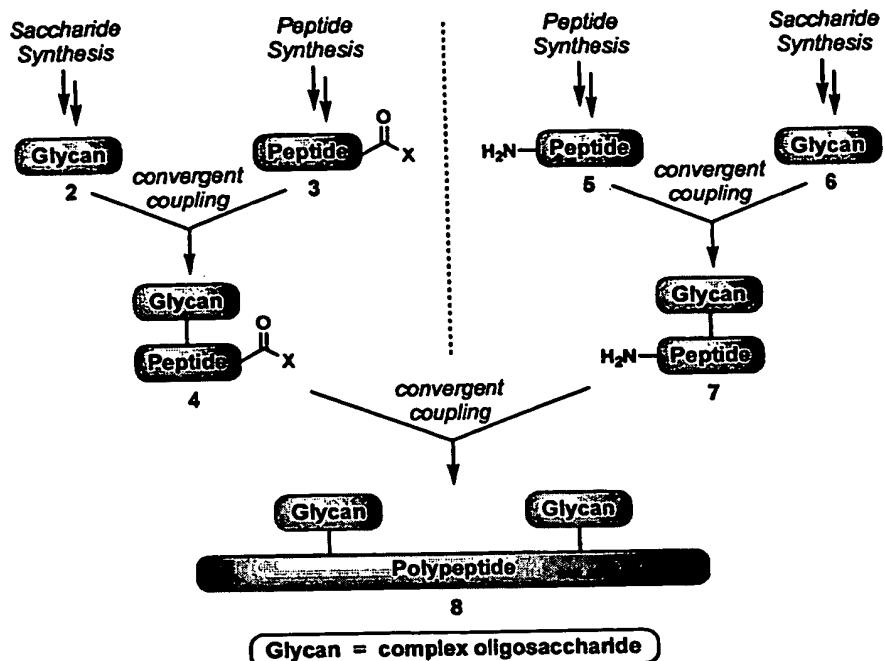
60/500,708; and International Patent Application Nos.: PCT/ US03/38453 and PCT/ US03/38471; each of the above-referenced patent documents are hereby incorporated by reference herein.

[0103] *Overview*

[0104] Following extensive earlier studies, wherein our chemistry was interfaced with important findings from other laboratories,¹² we demonstrated the ability to ligate fully synthetic N-linked glycopeptides containing various complex oligosaccharides to larger polypeptide domains.¹³ For example, major glycosylated subsections of prostate specific antigen (PSA)^{Error! Reference source not found.a} and gp120¹⁴ could be obtained. Provided herein is an exemplary convergent approach to access polypeptides multiply glycosylated at designated sites.¹⁵ As discussed previously, glycoproteins have potentially important clinical roles in the contexts of vaccines,² diagnostics,³ and therapeutics.⁴ The method of the present invention thus has important applications in the field of medicine since it offers access to complex therapeutically useful glycoproteins.

[0105] Scheme 7 schematizes an embodiment of the present invention. For example, glycopeptides 4 and 7 may be generated in a convergent way from their respective glycan and peptide precursors, and may be coupled with full convergence to reach 8.

[0106] **Scheme 7.** A convergent synthesis of bifunctional glycopeptides.



[0107] In one embodiment, the synthesis of a suitable glycopeptide acyl donor for ligation with 7 was investigated. One approach was based on utilizing building block 4, in the form of a thioester, as acyl donor. However, the preparation of a thioester such as 4 by convergent means¹⁶ appeared to present difficulties.

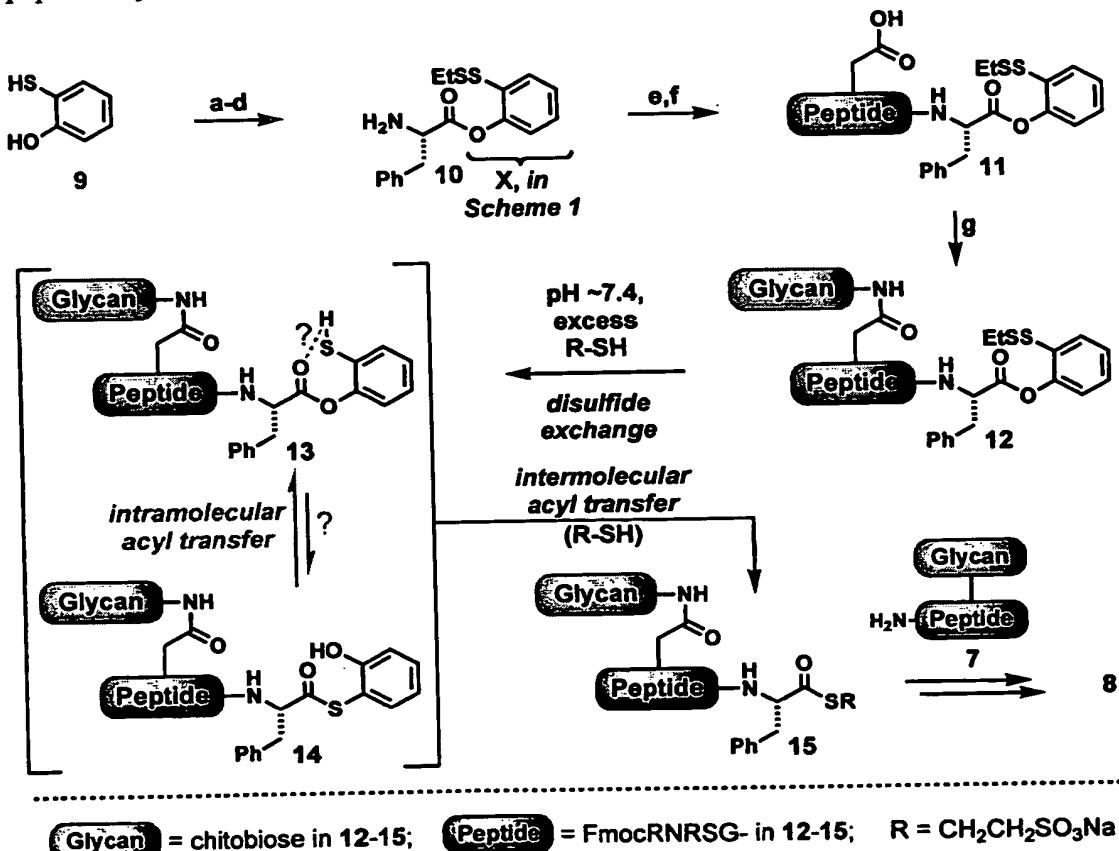
[0108] It was in pondering this problem that an attractive possibility presented itself (Scheme 8). The synthesis of a phenolic ester equipped with an unsymmetrical disulfide (*cf.* 10) was then investigated. The phenolic ester linkage would in itself probably not manifest sufficient reactivity to serve as a viable acylating agent. However, equipped as it would be with an *ortho*-disulfide moiety, the phenolic ester could operate as an incipient acyl donor. Thus, reduction of the disulfide in 12, as shown, might well set the stage for *in situ* elaboration into a thioester (*cf.* 13→14) by O→S acyl migration. Thioester 14, present in an unfavorable, but dynamic equilibrium, could perhaps be interdicted by the machinery appropriate to native chemical ligation (NCL),^{12c} ultimately culminating in 8.

[0109] While the proposal for the intermediacy of a discreet thioester 14 is certainly attractive, it is not necessarily an obligatory event en route to ligation. Thus, as suggested in structure 13, the presence of a free *ortho*-benzene thiol function might well enhance the acylating facility of the phenolic ester,¹⁷ perhaps through *in situ* hydrogen bonding or anchimeric

assistance. In either case, the sequence is set into motion only upon reductive cleavage of the disulfide.

[0110] The synthesis of the masked glycopeptide acyl donor was accomplished as follows (Scheme 8). Commercially available 2-mercaptophenol (9) was oxidized to its symmetrical disulfide. Subsequent exchange with excess ethyl disulfide¹⁸ furnished an unsymmetrical, aryl-alkyl disulfide. Acylation of the phenolic hydroxyl group with Boc-phenylalanine followed by acidic cleavage of the Boc protecting group afforded phenylalanine derivative 10. Standard peptide coupling of 10 with a fully protected peptide acid¹⁹ followed by global deprotection provided 11,²⁰ which contains a free aspartate residue. Aspartylation of a chitobiose-derived glycosylamine with 11 afforded the virtual acyl donor 12, corresponding to our target structure 4 (Scheme 7).

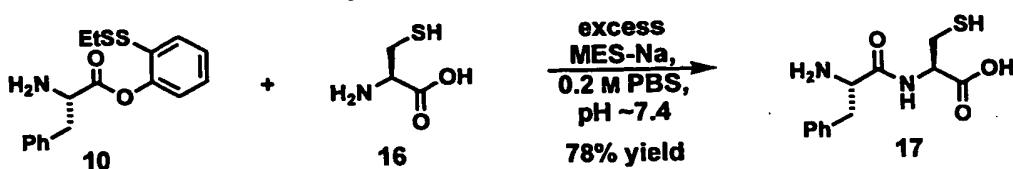
[0111] Scheme 8. A structure, synthesis, and proposed mechanism of a masked glycopeptide acyl donor.^a



[0112] ^a Reagents and conditions: (a) I₂, MeOH, H₂O; (b) BF₃•OEt₂, EtSSEt, CH₂Cl₂, 99%, 2 steps; (c) Boc-Phe-OH, EDCI, DMAP, CH₂Cl₂/THF, 93%; (d) 4N HCl/dioxane, 94%; (e) Fmoc-Arg(Pbf)-Asp('Bu)-Arg(Pbf)-Ser('Bu)-Gly-OH, HATU, DIEA, DMF, 61%; (f) TFA:phenol:Et₃SiH:H₂O 35:2:1:1, 60%; (g) GlcNAc β 1→4GlcNAc β -NH₂, HATU, DIEA, DMSO, 52%.

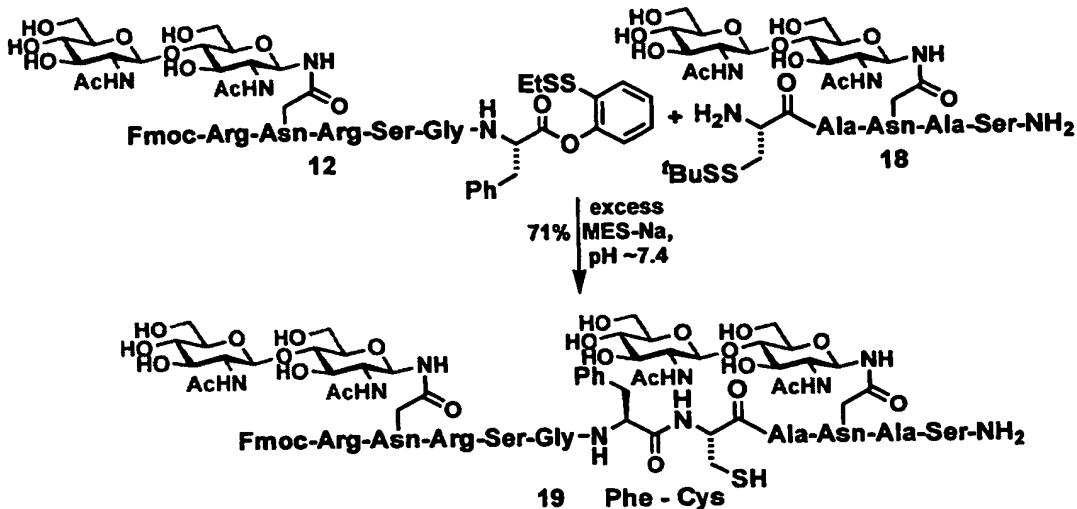
[0113] As a preliminary probe of the practicability of this scheme, phenylalanine derivative **10** was subjected to the neutral, aqueous, reducing reaction conditions described above, along with free cysteine (Scheme 9). The desired free dipeptide Phe-Cys (**17**) was obtained in 78% isolated yield. Only a single product was observed by LCMS and ¹H NMR, strongly suggesting that epimerization at the α -carbon of phenylalanine did not occur.

[0114] Scheme 9. An example of the latent acyl donor function.



[0115] The synthesis of bifunctional glycopeptides was then carried out. Thus, **12** was treated with excess sodium 2-mercaptoethanesulfonate (MES-Na) at neutral pH in aqueous phosphate buffered saline (PBS) in the presence of **18**,²¹ containing a latent N-terminal cysteine residue (Scheme 10). As observed by LCMS during the reaction, **12** was almost instantaneously (less than 3 min) converted to the thioester derived from MES-Na (*cf.* **15**); very little hydrolysis to the free carboxylic acid was noted. Over the course of the next several hours, we observed a gradual decrease in the amount of this thioester, with a concomitant increase in the desired product **19**. Only one compound of the appropriate mass was observed by LCMS and ¹H NMR, again indicating a lack of epimerization.

[0116] Scheme 10. Exemplary convergent coupling of functionalized glycopeptides.



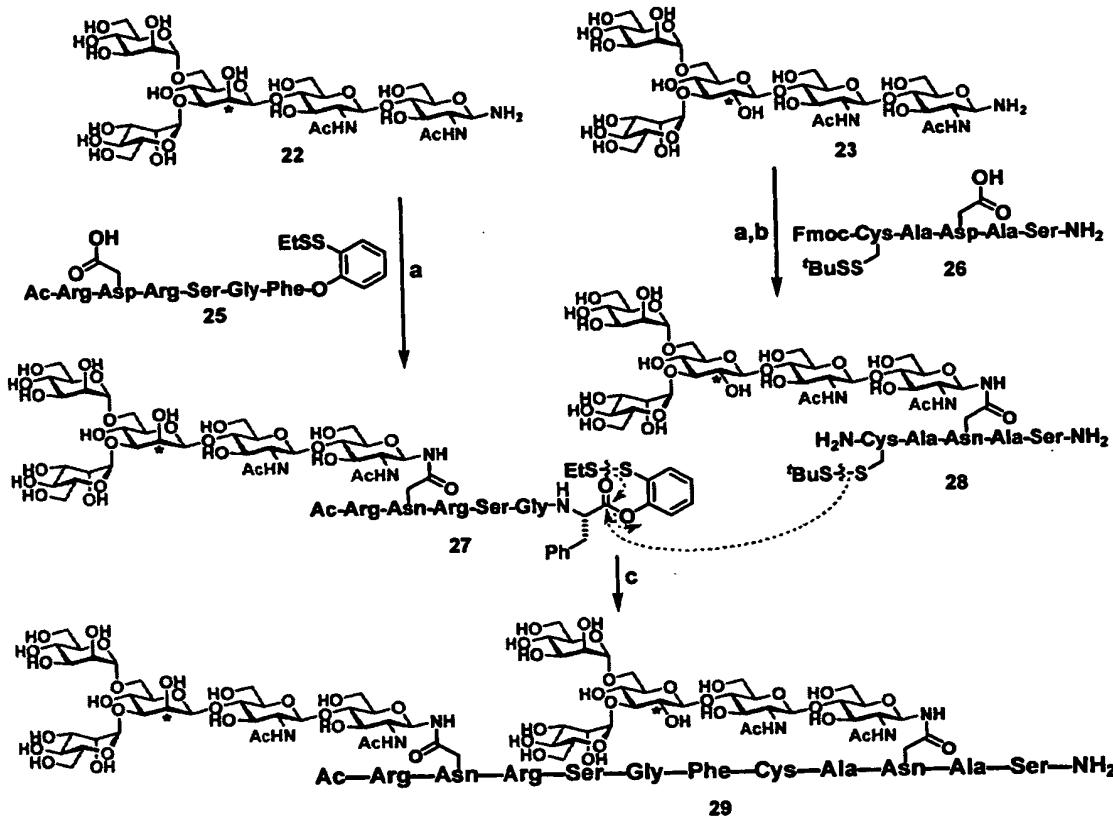
[0117] With this proof of principle in hand, attention was directed to investigating the generality of the method. For example, applicability of the method to incorporation of *O*-linked domains²² was established, as shown through the synthesis of 20. Notably, the histidine-containing system 21 was also assembled (for structure of 21 and 20, see Figure 1). Since activated C-terminal histidine residues are known to be particularly susceptible to epimerization,²³ a prophylactic dinitrophenyl (DNP) protecting group²⁴ was used for the imidazole τ -nitrogen during the synthesis. As expected, the DNP group was cleaved concurrently during the mildly nucleophilic coupling reaction. No indications of epimerization were observed by LCMS or ^1H NMR.

[0118] Validation of the highly convergent methodology in the context of complex glycans was then investigated. Syntheses of glycosylamines 22¹³ and 23²⁵ (Figure 2) have been reported previously. It will be noted that these compounds differ in a single stereogenic locus in the interior C-ring (see asterisks). Thus 22 corresponds to a fully stereochemically competent *N*-linked high mannose oligosaccharide. In contrast, its C-ring C-2 epimer is an unnatural stereochemical mutant. Studies aimed at evaluating whether high-mannose binding lectins can discriminate between point mutants that differ in only one out of a total of 25 stereochemical loci are underway.

[0119] Preparation of pentasaccharide-derived glycopeptide coupling partners proceeded via Lansbury aspartylation^{12b} of 22 and 23 with 25 and 26, which afforded 27 and 28,

respectively. Coupling of **28** with a relatively simple chitobiose-containing acyl donor gave **24** (Figure 2) in 77% yield.

[0120] **Scheme 11.** Convergent coupling of two functionalized glycopeptides containing distinct, structurally complex glycans.^a



[0121] ^a Reagents and conditions: (a) **25** or **26**, HATU, DIEA, DMSO, 50% for **27** (from **22**), 71% for **28** (from **23**); (b) 20% piperidine in DMF, 63%; (c) 0.2 M phosphate, 0.2 M NaCl, pH ~7.4, excess MES-Na, 75%.

[0122] Coupling of **27** and **28** was accomplished smoothly upon reductive cleavage of the disulfide linkage in **27**, affording **29** as shown in Scheme 11. Aside from validating the methodology in a striking way, the synthesis of **29** serves to pinpoint the power of the total synthesis approach: Since one of its carbohydrates is unnatural, **29** cannot readily be obtained from natural sources or via enzymatic manipulations.

[0123] In summary, a convergent method for the synthesis of diglycosylated peptides is described herein. The mechanistic rationale set forth in Scheme 8 is supported by the

identification of the MES-Na thioester as a reaction intermediate. Furthermore, potential hydrolysis and C-terminal epimerization of the glycopeptide acyl donor have been suppressed. It seems likely that the method and logic set forth above will enjoy application in the building of complex glycopeptides of biological and even medicinal consequence.

[0124] It will be appreciated that the method can readily be applied to the preparation of multi-glycosylated peptides (*e.g.*, comprising more than two non-adjacent glycosides covalently linked at designated sites).

[0125] **General Reaction Procedures:**

[0126] Unless mentioned specifically, reaction mixtures were stirred using a magnetically driven stirrer bar. Reactions involving air or moisture-sensitive reagents or intermediates were performed under argon or nitrogen atmosphere in glassware which had been heat gun or flame-dried under high vacuum. An inert atmosphere refers to either dry argon or dry nitrogen. Reactions were monitored either by thin layer chromatography, by proton nuclear magnetic resonance (NMR) or by high-pressure liquid chromatography (HPLC), of a suitably worked up sample of the reaction mixture.

[0127] Indicated reaction temperatures refer to those of the reaction bath, while room temperature (rt) is noted as 22 °C. Preparative reactions were stirred magnetically. Tetrahydrofuran (THF), diethyl ether (Et₂O), methylene chloride (CH₂Cl₂), and toluene were obtained from a dry solvent system (activated alumina columns, positive pressure of argon). All other solvents were used as received in Sure/Seal bottles (Aldrich). Triethylamine (Et₃N), diisopropylethylamine (*i*-Pr₂NEt), pyridine, 2,6-lutidine, and chlorotrimethylsilane (TMSCl) were distilled from CaH₂ immediately prior to use. All other reagents were purchased from Aldrich at the highest commercial quality and used without further purification, with the exception of the Stryker reagent which was purchased from Fluka, the RCM catalysts 16 and 17 which were purchased from Strem, and biotin-dPEG₄-hydrazide which was purchased from Quanta Biodesign.

[0128] **General Work Up Procedures:**

[0129] Unless mentioned specifically, reaction mixtures were cooled to room temperature or below then quenched, when necessary, with either water or a saturated aqueous solution of ammonium chloride. Desired products were extracted by partitioning between water

and a suitable water-immiscible solvent (e.g. ethyl acetate, dichloromethane, diethyl ether). The desired product containing extracts were washed appropriately with water followed by a saturated solution of brine. On occasions where the product containing extract was deemed to contain residual oxidants, the extract was washed with a 10% solution of sodium sulphite in saturated aqueous sodium bicarbonate solution, prior to the aforementioned washing procedure. On occasions where the product containing extract was deemed to contain residual acids, the extract was washed with saturated aqueous sodium bicarbonate solution, prior to the aforementioned washing procedure (except in those cases where the desired product itself had acidic character). On occasions where the product containing extract was deemed to contain residual bases, the extract was washed with 10% aqueous citric acid solution, prior to the aforementioned washing procedure (except in those cases where the desired product itself had basic character). Post washing, the desired product containing extracts were dried over anhydrous magnesium sulphate, and then filtered. The crude products were then isolated by removal of solvent(s) by rotary evaporation under reduced pressure, at an appropriate temperature (generally less than 45°C).

[0130] **General Purification Procedures:**

[0131] Unless mentioned specifically, chromatographic purification refers to flash column chromatography on silica, using a single solvent or mixed solvent as eluent. Suitably purified desired product containing elutes were combined and concentrated under reduced pressure at an appropriate temperature (generally less than 45°C) to constant mass. Final compounds were dissolved in 50% aqueous acetonitrile, filtered and transferred to vials, then freeze-dried under high vacuum before submission for biological testing.

[0132] **Analytical Equipment:**

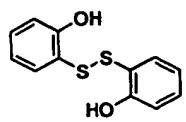
[0133] **HPLC:** All separations involved a mobile phase of 0.05% TFA (v/v) in water (solvent A)/0.0425% TFA in acetonitrile (solvent B). Preparative, semipreparative, and analytical HPLC separations were performed using a Rainin HXPL solvent delivery system equipped with a Rainin UV-1 detector and one of the following Dynamax-60Å C18 axial compression columns 250 mm in length equipped with a similarly packed guard column: 41.4 mm diameter (prep), 21.4 m diameter (semiprep), or 4.6 mm diameter (analytical). Separations were performed at flow rates of 48 mL/min (prep), 16 mL/min (semiprep), or 1 mL/min (analytical), and were monitored at a wavelength between 214 and 230 nm, depending on

column loading. LCMS chromatographic separations were performed using a Waters 2695 Separations Module and a Waters 996 Photodiode Array Detector equipped with a Varian Microsorb C18 2 x 150 mm column at a flow rate of 0.2 mL/min.

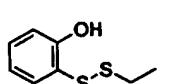
[0134] **ESMS and LCMS:** Electrospray mass spectroscopy and LCMS analyses were obtained on a Waters Micromass ZQ mass spectrometer in conjunction with the Waters HPLC apparatus described above.

[0135] **NMR:** ^1H and ^{13}C NMR spectra were recorded on Bruker instruments in CDCl_3 , CD_3OD or D_2O at 400 or 500 MHz for ^1H and 100 or 125 MHz for ^{13}C .

[0136] **Reagents:** All commercial materials were used as received unless otherwise noted. The following solvents were obtained from a dry solvent system and used without further purification: THF, diethyl ether, toluene, and DCM. Reagents were obtained from Aldrich or as noted, with the following exceptions: amino acids and resins for solid phase peptide synthesis were purchased from NovaBiochem; Biosynthesis grade DMF from EM Science; and all other solvents from Fisher Scientific (HPLC grade).

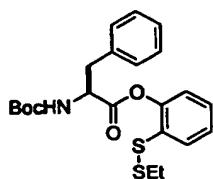


[0137] To a stirred, biphasic solution of 2-mercaptophenol (1.0 g, 7.9 mmol) in H_2O (5.1 mL) was added, drop wise, a solution of iodine (1.0 g 4.0 mmol) in methanol (3.5 mL). When the brown iodine color persisted the solution was diluted with ethyl acetate and water. The aqueous layer was removed and extracted with an additional portion of ethyl acetate. The combined organic layers were dried and washed with brine then dried (Na_2SO_4) and concentrated to give a brown oil which was used without purification (1.5 g). The product still contains iodine. ^1H NMR (CDCl_3 , 500 MHz): δ 7.33-7.37 (m, 2H), 7.22-7.24 (m, 2H), 6.99-7.01 (m, 2H), 6.82-6.85 (m, 2H), 6.22 (brs, 2H). ^1H NMR (CDCl_3 , 400 MHz): δ 7.35 (dt, $J = 8.16, 1.65$ Hz, 2H), 7.22 (dd, $J = 7.52, 1.65$ Hz, 2H), 7.00 (dd, $J = 8.16, 1.15$ Hz, 2H), 6.22 (s, 2H) ^{13}C NMR (CDCl_3 , 125 MHz): δ 157.3, 136.6, 133.6, 121.4, 120.3, 116.1. ESI-MS: Calcd. for $\text{C}_{12}\text{H}_{10}\text{O}_2\text{S}_2$ [$\text{M}+\text{NH}_4$] $^+$ 267.8 Found: 267.8.



[0138] To a stirred solution of the disulfide (1.5 g crude, ~4 mmol) in CH_2Cl_2 (25 mL) was added ethyldisulfide (10.5 mL, 79.9 mmol) and then $\text{BF}_3\text{-OEt}_2$ (10.1 mL, 79.9 mmol). The reaction was stirred at room temperature for three hours and then carefully quenched by the addition of (aq.) NaHCO_3 . The organic layer was drained

and the aqueous layer was extracted with an additional portion of CH_2Cl_2 . The combined organic layers were dried (MgSO_4) and concentrated to give a yellow oil. Purification by silica gel chromatography (20% ethyl acetate in hexane) gave the desired product as a clear, slightly yellow oil (1.45 g, 99%). ^1H NMR (CDCl_3 , 500 MHz): δ 7.48-7.51 (m, 1H), 7.28-7.32 (m, 1H), 6.99-7.01 (m, 1H), 6.86-6.9 (m, 1H), 6.34 (brs, 1H), 2.78 (q, $J = 7.4$ Hz, 2H), 1.35 (t, $J = 7.4$ Hz, 3 H). ^{13}C NMR (CDCl_3 , 125 MHz): δ 156.9, 135.2, 132.2, 121.0, 116.2, 32.4, 14.1. ESI-MS: Calcd. for $\text{C}_8\text{H}_{10}\text{OS}_2$ [M+Na] $^+$ 208.8 Found: 208.8.

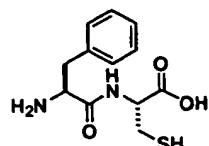


[0139] To a solution of the phenol (1.45 g, 8 mmol) and Boc-Phe-OH (2.65 g, 10 mmol) in CH_2Cl_2 (25 mL) and THF (5 mL) was added EDCI (1.92 g, 10 mmol) and DMAP (98 mg, 0.8 mmol). The resulting solution was stirred at room temperature for 18 hr at which point the volatile materials were removed *in vacuo*. The resulting oil was taken up in EtOAc and washed with 1N HCl, H_2O , and then brine. The organic layer was dried (Na_2SO_4) and concentrated to give a slightly yellow oil. Purification by silica gel chromatography (30% ethyl acetate in hexane) gave a clear, colorless oil (3.5 g, >99%). ^1H NMR (CDCl_3 , 400 MHz): δ 7.78-7.81 (m 1H), 7.23-7.36 (m, 7H), 7.01-7.03 (m, 1H), 5.02 (d, $J = 8.16$ Hz, 1H), 4.86-4.91 (m, 1H), 3.31 (abx, $J_{ab} = 13.98$ Hz, $J_{ax} = 5.41$ Hz, $J_{bx} = 7.22$ Hz, 2H), 2.72 (q, $J = 7.33$ Hz, 2H), 1.42 (s, 9H), 1.29 (t, $J = 7.33$ Hz, 3H). ESI-MS: Calcd. for $\text{C}_{22}\text{H}_{27}\text{NO}_4\text{S}_2$ [M+Na] $^+$ 456.0 Found: 456.0.



[0140] To a 250 mL round-bottomed flask equipped with a magnetic stir bar was added the above phenylalanine derivative (3.0 g, 6.9 mmol) as well as a 4M solution of HCl in dioxane (86 mL, 345 mmol) and the reaction was stirred under argon at room temperature for 1.5 hr. At that point the reaction was concentrated *in vacuo*, leaving a white solid. This material was triturated with ether and subsequently concentrated. This process was repeated (3x) leaving the crude product as a white solid. The material was dissolved in 30% B, shell frozen and lyophilized to give a white powder (2.17 g, 94%). ^1H NMR (CD_3OD , 400 MHz): δ 7.93-7.95 (m, 1H), 7.52-7.42 (m, 7H), 7.23-7.21 (m, 1H), 4.81 (dd, $J = 8.35, 5.73$ Hz, 1H), 3.56 (abx, $J_{ab} = 14.48$ Hz, $J_{ax} = 5.73$ Hz, $J_{bx} = 8.35$ Hz, 2H), 2.85 (q, $J = 7.34$, 2H), 1.37 (t, $J = 7.34$, 3H). ^{13}C NMR (CDCl_3 , 125 MHz): δ 169.0, 149.4,

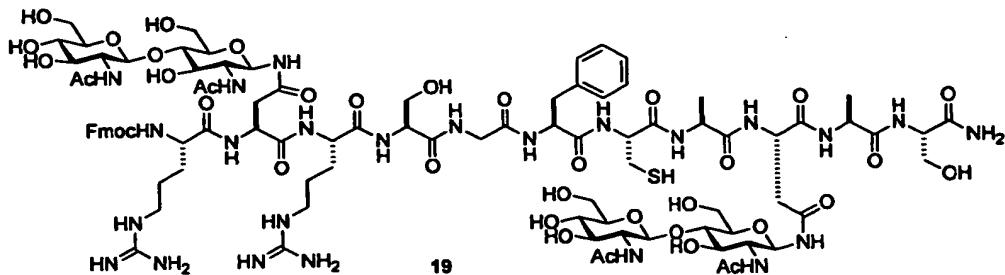
135.6, 131.9, 131.3, 131.0, 130.7, 129.9, 129.6, 129.1, 123.9, 55.6, 38.0, 34.1, 14.9. ESI-MS:
Calcd. for C₁₇H₁₉NO₂S₂ [M+H]⁺ 334.1 Found: 334.2.



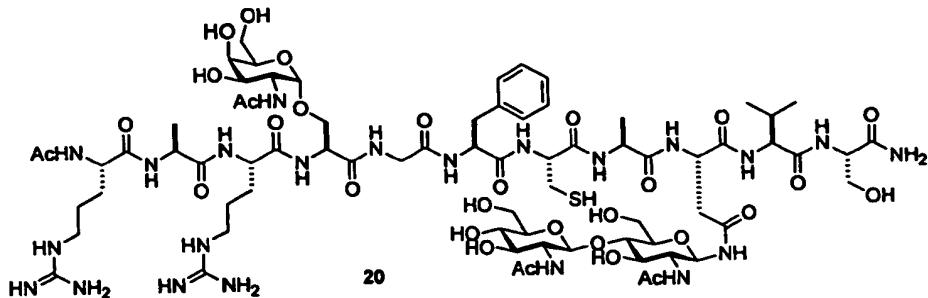
[0141] The phenylalanine derivative **10** (15 mg, 45 μ mol) and L-cysteine (6 mg, 49 μ mol) were placed into a LCMS vial along with a flea-sized stirbar. In a second vial were mixed MES-Na (25 mg, 150 μ mol) and phosphate buffered saline (PBS) (0.2M NaCl, 0.2M phosphate, pH = 7.5, 2 mL). The MES-Na solution was then added directly to the amino acids, and the reaction was monitored by LCMS. After two hours the reaction appeared to be complete and TCEP (129 mg, 450 μ mol) was added. This was stirred for 1 hour and then injected directly onto the HPLC for purification. The desired compound was obtained as a white powder after lyophilization (9.4 mg, 78%). ¹H NMR (D₂O): δ 7.36-7.45 (m, 3H), 7.31-7.33 (m, 2H), 4.63 (dd, *J* = 6.61, 4.83 Hz, 1H), 4.34 (dd, *J* = 7.24, 7.18 Hz, 1H), 3.25 (abx, *J_{ab}* = 14.1 Hz, *J_{ax}* = 7.18 Hz, *J_{bx}* = 7.24 Hz, 2H), 2.96 (abx, *J_{ab}* = 14.2 Hz, *J_{ax}* = 4.83 Hz, *J_{bx}* = 6.61 Hz, 2H). LCMS: 5-65%B over 20 min, rt = 8.53 min. HPLC: 5-95%B over 30 min, rt = 11.45 min. ESI-MS: Calcd. for C₁₂H₁₆N₂O₃S [M+H]⁺ 269.1, Found: 269.1

[0142] Exemplary ligation conditions:

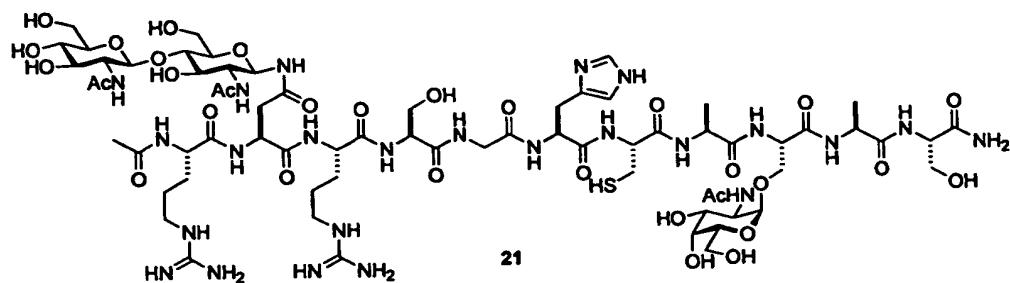
[0143] The two glycopeptide halves (**12**, 2.2 mg, 1.44 μ mol) (**18**, 1.4 mg, 1.44 μ mol) were placed in a LCMS vial along with a flea-sized stirbar. A stock solution of MESNa (18.3 mg, 111 mmol) in phosphate buffered saline (0.2M NaCl, 0.2M phosphate, pH = 7.4, 1 mL) was made and of this, 600 μ L was added to the glycopeptides. The reaction was monitored by LCMS and, once finished, TCEP (25 mg, 0.087 mmol) was added and the solution stirred for 2 hr then injected directly onto the HPLC for purification.



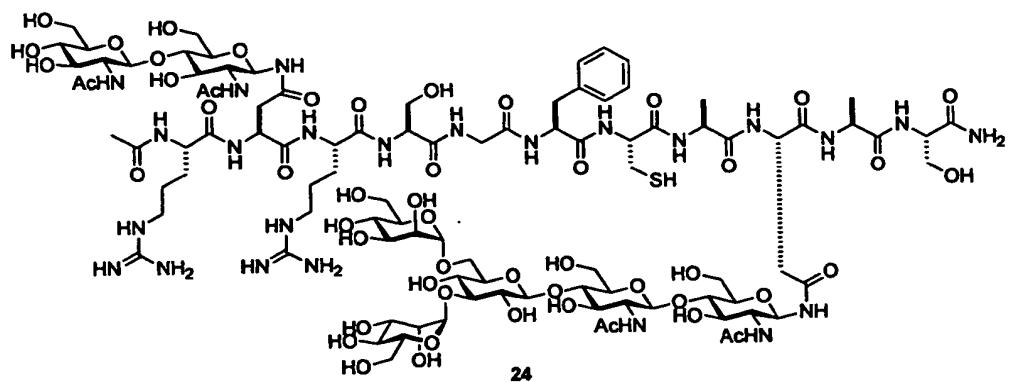
[0144] LCMS: 5-65%B over 20 min, rt = 9.98 min. HPLC: 25-55%B over 30 min, rt = 7.65 min. ESI-MS: Calcd. for $C_{93}H_{138}N_{24}O_{37}S$ [M+2H]²⁺ 1108.5, Found: 1108.6, [M+3H]³⁺ 739.3, Found: 739.5.



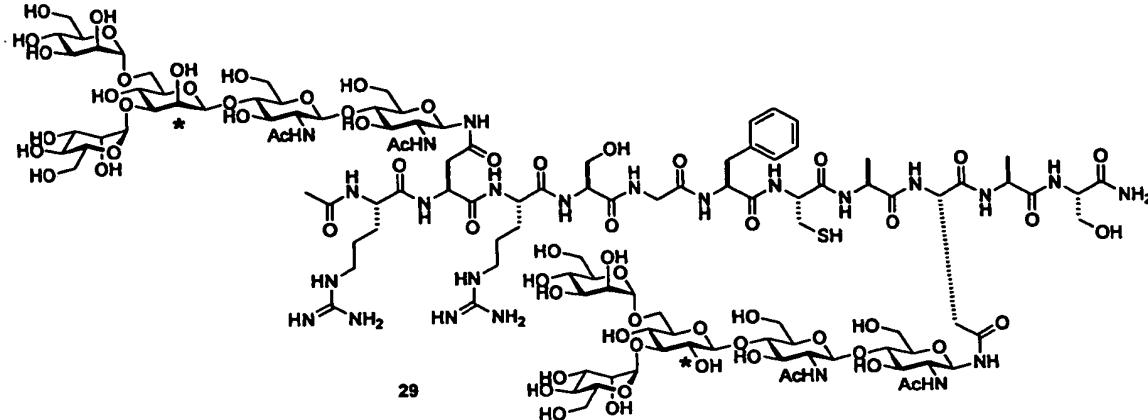
[0145] LCMS: 5-65%B over 20 min, rt = 11.40 min. HPLC: 5-65%B over 20 min, rt = 9.85 min. ESI-MS: Calcd. for $C_{73}H_{120}N_{22}O_{30}S$ [M+2H]²⁺ 909.4, Found: 909.5, [M+3H]³⁺ 606.6, Found: 606.8.



[0146] LCMS: 5-65%B over 20 min, rt = 4.02 min. HPLC: 5-65%B over 20 min, rt = 6.71 min. ESI-MS: Calcd. for $C_{73}H_{120}N_{22}O_{30}S$ [M+2H]²⁺ 898.4, Found: 898.6.



[0147] LCMS: 5-65% B over 20 min, rt = 7.81 min. HPLC: 5-65% B over 20 min, rt = 8.39 min. ESI-MS: Calcd. for $C_{98}H_{160}N_{24}O_{51}S$ [M+2H]²⁺ 1261.5, Found: 1261.5, [M+3H]³⁺ 841.4, Found: 841.5.



[0148] LCMS: 5-45% B over 20 min, rt = 10.82 min. HPLC: 5-45% B over 20 min, rt = 9.31 min. ESI-MS: Calcd. for $C_{98}H_{160}N_{24}O_{51}S$ [M+2H]²⁺ 1504.6, Found: 1504.9, [M+3H]³⁺ 1003.4, Found: 1003.7.

ABBREVIATIONS AND GLOSSARY

- [0149] A: alanine
- [0150] Ac: acetyl
- [0151] ACT: α 1-antichymotrypsin
- [0152] Ala: alanine
- [0153] Arg: arginine
- [0154] Asn: asparagine
- [0155] Asp: aspartic acid
- [0156] Bn: benzyl
- [0157] Boc: *tert*-butyloxycarbonyl
- [0158] BPH: benign prostatic hyperplasia
- [0159] BSP: benzenesulfinyl piperidine
- [0160] Bu: butyl
- [0161] Bz: benzoyl
- [0162] CAN: ceric ammonium nitrate

[0163]	coll: <i>sym</i> -collidine
[0164]	C-terminus: peptide carbonyl terminus
[0165]	Cys: cysteine
[0166]	D: aspartic acid
[0167]	DIEA: <i>N,N</i> -diisopropylethylamine
[0168]	DMAP: <i>N,N</i> -Dimethylaminopyridine
[0169]	DMF: dimethyl formamide
[0170]	DMSO: dimethyl sulfoxide
[0171]	DTBMP: di- <i>tert</i> -butylmethylpyridine
[0172]	DTBP: di- <i>tert</i> -butylpyridine
[0173]	Et: ethyl
[0174]	Fmoc: 9-fluorenylmethyloxycarbonyl
[0175]	G: glycine
[0176]	Gal: galactose
[0177]	Glc: glucose
[0178]	Gln: glutamine
[0179]	Glu: glutamic acid
[0180]	Gly: glycine
[0181]	H: histidine
[0182]	HATU: 7-azahydroxybenzotriazolyl tetramethyluronium hexafluorophosphate
[0183]	His: histidine
[0184]	Ile: isoleucine
[0185]	K: lysine
[0186]	kDa: kilodaltons
[0187]	KLH: keyhole limpet hemocyanin
[0188]	L: leucine
[0189]	Leu: leucine
[0190]	Lys: lysine
[0191]	Man: mannose
[0192]	MES-Na: 2-mercaptopethanesulfonic acid, sodium salt
[0193]	MHC: major histocompatibility complex

[0194]	N: asparagine
[0195]	NAc: <i>N</i> -acetyl
[0196]	NCL: native chemical ligation
[0197]	N-terminus: peptide amine terminus
[0198]	<i>O</i> -linked: linked through an ethereal oxygen
[0199]	Pam3Cys: tripalmitoyl-S-glycerylcysteinylserine
[0200]	PBS: phosphate-buffered saline
[0201]	Ph: phenyl
[0202]	Phth: phthalimido-
[0203]	PMB: <i>p</i> -methoxybenzyl
[0204]	Pro: proline
[0205]	GP120: prostate specific antigen
[0206]	Py: pyridine
[0207]	QS21: a glycosidic immunoadjuvant
[0208]	R: arginine
[0209]	S: serine
[0210]	sat. aq.: saturated aqueous
[0211]	Ser: serine
[0212]	T: threonine
[0213]	TBAF: tetra- <i>n</i> -butylammonium fluoride
[0214]	TBS: <i>tert</i> -butyldimethylsilyl
[0215]	<i>t</i> Bu: <i>tert</i> -butyl
[0216]	Tf: trifluoromethanesulfonate
[0217]	TFA: Trifluoroacetic acid
[0218]	THF: tetrahydrofuran
[0219]	Thr: threonine
[0220]	t-GP120: total prostate specific antigen
[0221]	TMSCl: Trimethylsilyl chloride
[0222]	Trp: tryptophan
[0223]	V: valine
[0224]	Val: valine

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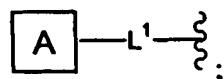
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- [0246] (21) Glycopeptide **18** was prepared as described previously (ref. 13).
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[0276] (51) There is an intermediate step in NCL during which there is a second transthio-esterification with MESNa. In our reactions we have never observed the free sulfhydryl of the phenyl ester. Rather complete conversion of the *N*-terminal to the corresponding MESNa thioester is observed within 3 min.

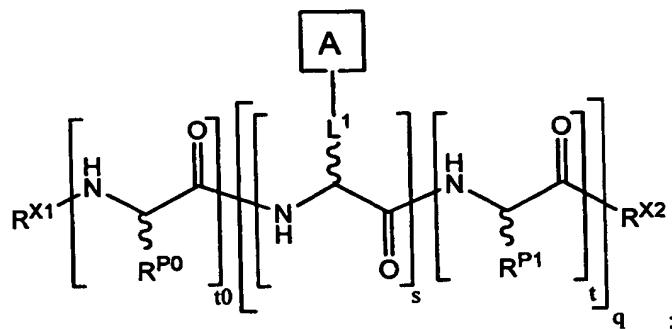
ABSTRACT

The present invention provides a method for preparing polyfunctionalized peptides and/or proteins at non-adjacent designated sites via native chemical ligation. In certain embodiments, the inventive method is a method for preparing a polyfunctionalized peptide comprising a peptidic backbone made up of four or more amino acids, wherein two or more non-adjacent amino acids are independently substituted with a moiety having the structure:



wherein A and L¹ are as defined herein.

In certain other embodiments, the inventive method allows the preparation of polyfunctionalized peptides having the general structure:



wherein A, R^{P0}, R^{P1}, R^{X1}, R^{X2}, L¹, t₀, s, t and q are as defined herein.

FIG. 1

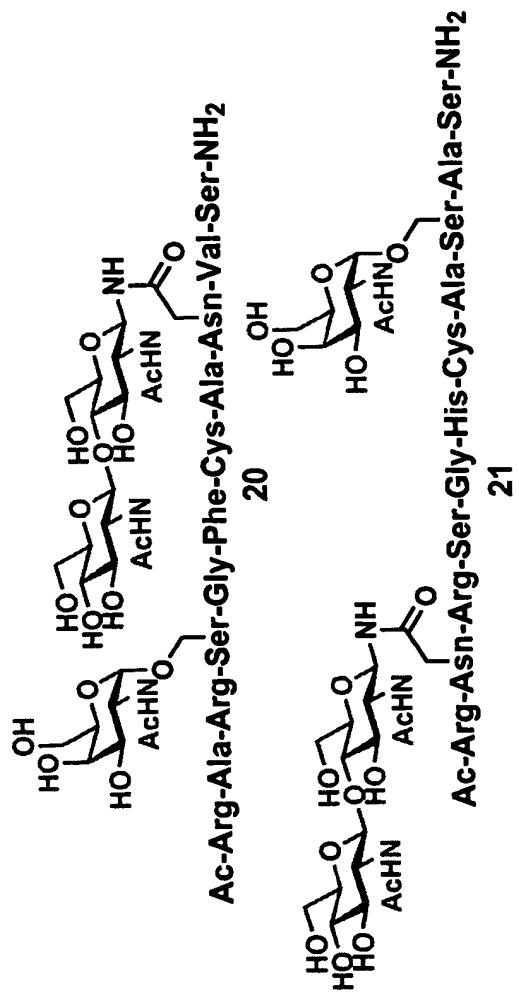
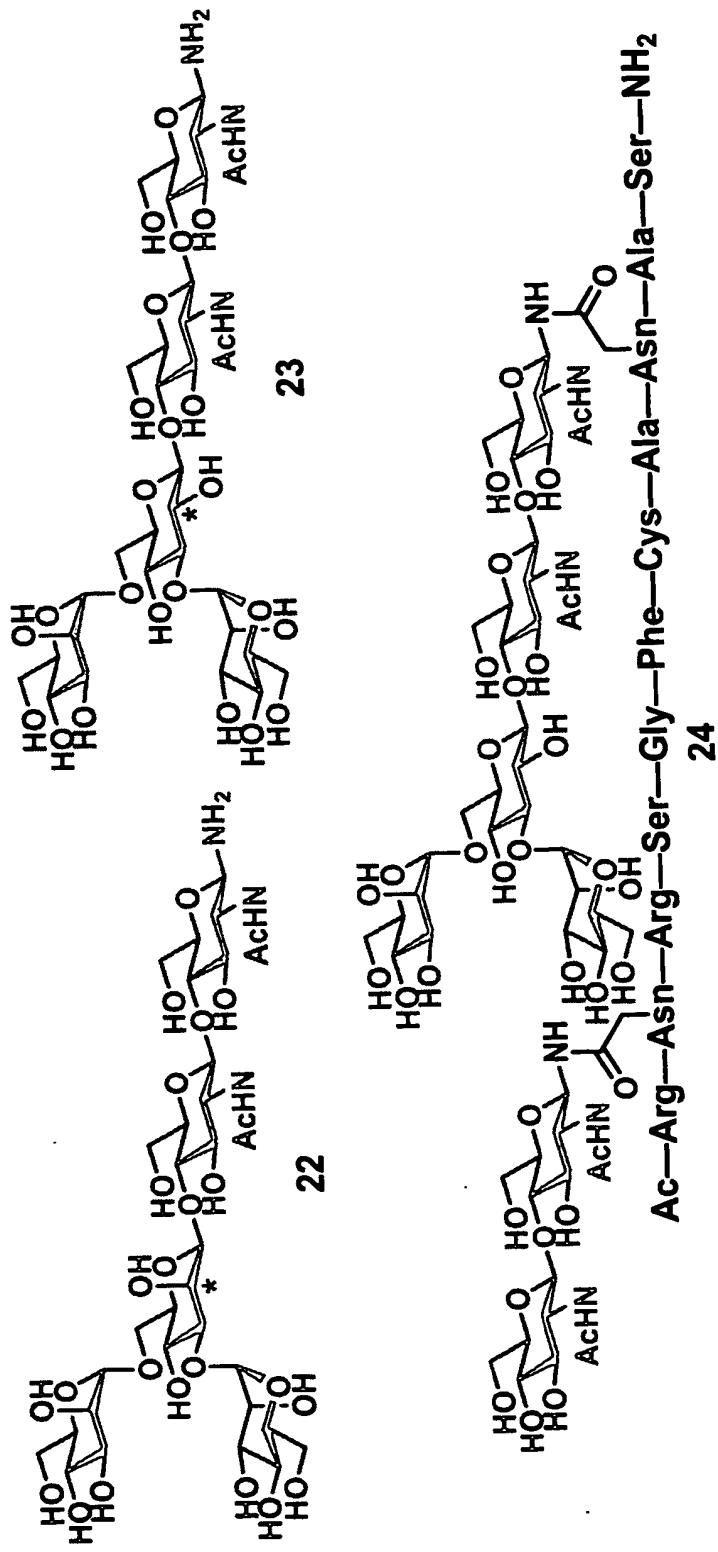


FIG. 2



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